

1

Lecture and Practice Proceedings & Objectives

- Have a flavor of the broadness of the drug design applications,
- Acquire the basic theoretical background,
- Practice the molecular graphics techniques,
- Know the free web-based tools developed at SIB,
- Use them for structure-based and ligand-based design

➔ You should be able to perform simple tasks of computer-aided drug design on whatever computer connected to the internet

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Lectures & Practices Agenda

Session	Lecture	Practice
1	Prologue: molecular representation	
	Introduction to (computer-aided) drug design	
	Origin of 3D structures	
	Molecular recognition	Use of UCSF chimera to analyze protein-ligand complexes
2	Binding free energy estimation	
	Introduction to molecular docking	Ligand-protein docking with AutoDock Vina
3	Introduction to molecular (virtual) screening	Ligand-based virtual screening with SwissSimilarity
4	Short introduction on target prediction of small molecules	Use of SwissTargetPrediction to perform reverse screening.
5	Introduction to ADME, pharmacokinetics, druglikeness	Estimate physicochemical, pharmacokinetic, druglike and related properties with SwissADME
6	Short introduction to bioisosterism	Use of SwissBioisostere to perform bioisosteric design

Installing UCSF ChimeraX

In this lecture/practical you will use the software UCSF ChimeraX for 3D structure visualisation and analysis.

This software is:

- free for teaching or academic research
- available for the most current platforms (Windows, Mac, Linux)
- open source (you can modify it for your needs if you know how to code in **python**. This is out of the scope of this lecture).

You can download the latest production release here:

<https://www.cgl.ucsf.edu/chimerax/download.html>

Please, install this software on your machine.

It will be mandatory for the practicals, but also useful for the theoretical lectures

[Download UCSF ChimeraX](#)  

ChimeraX is the state-of-the-art visualization program from the [Resource for Biocomputing, Visualization, and Informatics](#) at UC San Francisco. It is free for academic, government, nonprofit, and personal use; commercial users, please see [commercial licensing](#). Please cite [ChimeraX](#) in publications.

See also:

- [ChimeraX Documentation](#)
- [System Requirements](#)
- [Change Log](#)
- [Download & Citation Counts](#)
- [Older Releases](#)
- [Common Problems](#)

Current releases:

- [Release Candidate Builds](#)
- [Production Builds](#)
- [Daily Builds](#)

ChimeraX 1.10 Release Candidate (15 June 2025)

Please try these candidates for the next production release. See the [change log](#) for a list of improvements since the last production release. If needed, new candidate releases with bug fixes are made before the production release is made. If your work depends on a build provided by the [ChimeraX Toolkit](#), you may want to defer updating until the toolkit has also been updated to work with this version.

Operating System	Distribution	Date	Notes
macOS	chimerax-candidate.dmg	15 Jun 2025	Includes native versions for M2, M1 and Intel Macs. Works on macOS 11 and newer. ► More Info...

► [Other releases](#)

ChimeraX 1.9

Production releases are stable versions for ChimeraX Toolkit bundles to work with. You may need to use an [older release](#) if a bundle you wish to use has not been updated yet. Showing releases for Mac. Due to the nature of the [ChimeraX Toolkit](#), we support multiple versions that we support, even with ARM cores running macOS before 14 should use the 1.10 release candidate or 1.11 daily builds. Since we are close to a release, a 1.9.1 release is not planned at this time.

Operating System	Distribution	Date	Notes
macOS	ChimeraX-1.9.dmg	11 Dec 2024	built: undefined commit: 2024-12-11 19:11:19 UTC size: 517.7 MB sha512: 62b16b73238664cb094118b7967991 sha256: 886750e591b304e8f2c753a04c9e2d3b707c22587d7a7a1b5058ca32567326ac

► [Other releases](#)

Daily Build

Daily builds are generated automatically each night from the [development source code](#) (see the [change log](#)). While a given build may have unforeseen problems, these are often fixed by the next day. Showing releases for Mac.

Operating System	Distribution	Date	Notes
macOS	chimerax-daily.dmg	15 Jun 2025	Includes native versions for Apple Silicon and Intel Macs. Works on macOS 12 and newer. ► More Info...

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The dedicated web site

This teaching has been conceived to alternate theoretical lectures and practicals, so that you will:

- experiment yourself the visualisation and analysis of ligand-protein 3D structures
- get a flavor of different tools of computer-aided drug design

To facilitate the process, a web site has been especially conceived for this teaching. You can find it here:

<http://www.drug-design-teaching.ch>

1. This web site will indicate you **when to switch between lecture and practicals**. For instance, you will be able to make Session 1 exercices just after the lecture on molecular recognition
2. The **booklet of the practicals and the PDF of the lecture** can be downloaded from the web site too

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 Swiss Institute of
 Bioinformatics

Lecture
 The PDF file of the lecture's slides can be found [here](#) for Session 1.
 Prologue: molecular representations
 Introduction to (computer-aided) drug design
 Origin of 3D structures
 Molecular recognition

 Binding free energy estimation
 Introduction to molecular docking

 Introduction to molecular (virtual) screening

 Short introduction on target prediction of small molecules

 Introduction to ADME, pharmacokinetics, druglikeness

Practice
 The PDF file of the practice booklet can be found [here](#).

Practice session 1: Introduction to UCSF Chimera
 Exercise 1. UCSF Chimera basics (n - Loading a structure, moving, zooming, selecting, rendering, analysing and saving)
 Exercise 2. Using surfaces in UCSF Chimera
 Exercise 3. Using lighting and effects in UCSF Chimera
 Exercise 4. Loading several structures and aligning with the MultiAlign Viewer

Practice session 2: Ligand-protein docking
 Exercise 5. Docking of anti-inflammatory drug Celecoxib into protein COX2
 Exercise 6. Structure-based optimisation of COX2 inhibitors

Practice Session 3. Ligand-based virtual screening with SwissSimilarity

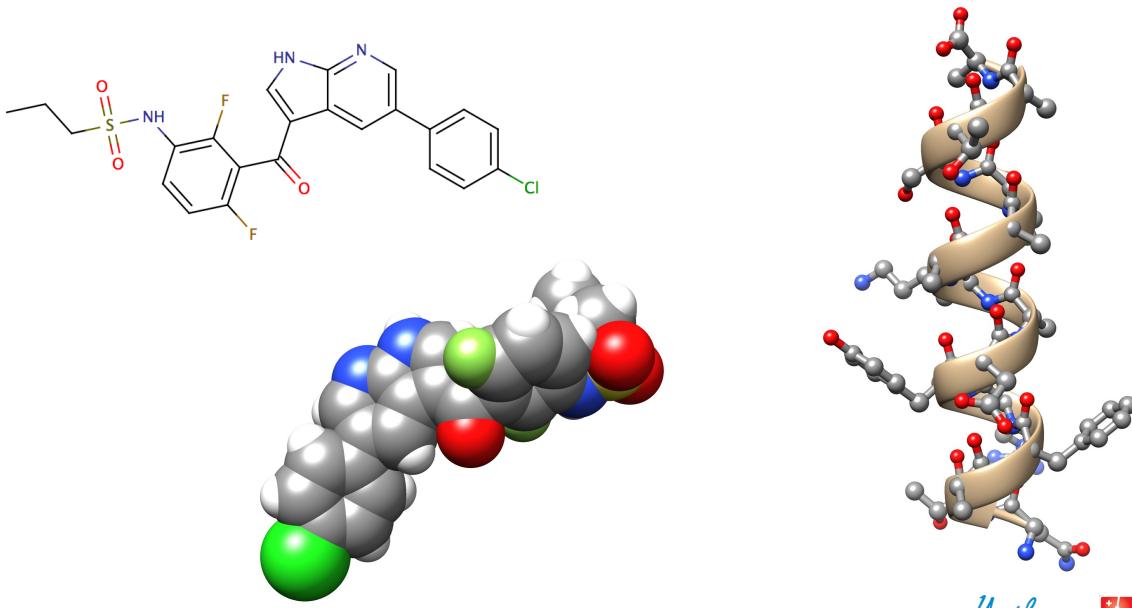
Practice Session 4. Reverse screening with SwissTargetPrediction

Links to download lecture and practice here

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Prologue: molecular representations



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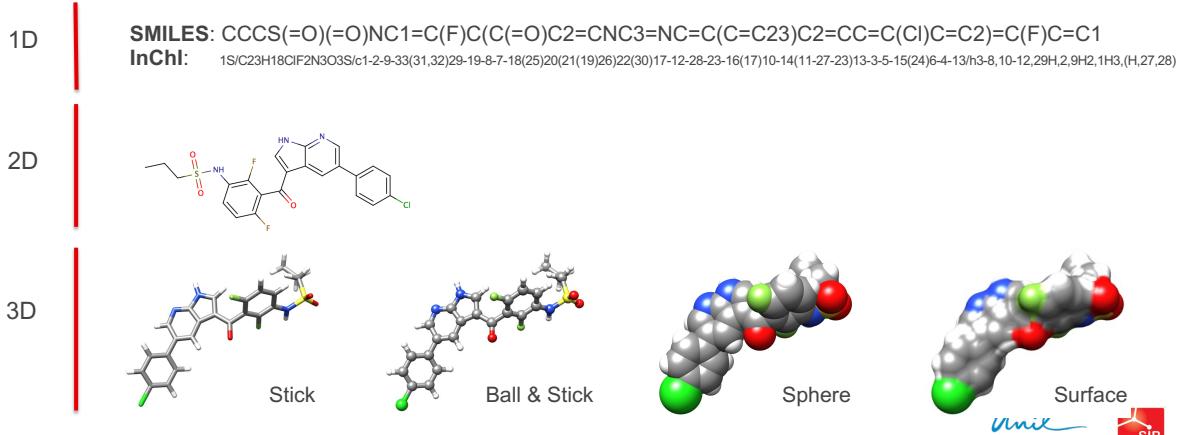
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Molecular representations – “small” molecules

Organic molecules of less than ~ 100 atoms are often referred to as “small” molecules, as opposed to biological macromolecules (i.e. proteins, DNA, etc.)

Small molecules can be represented in 1D, 2D or 3D:

Example of Vemurafenib (BRAF V600E inhibitor)



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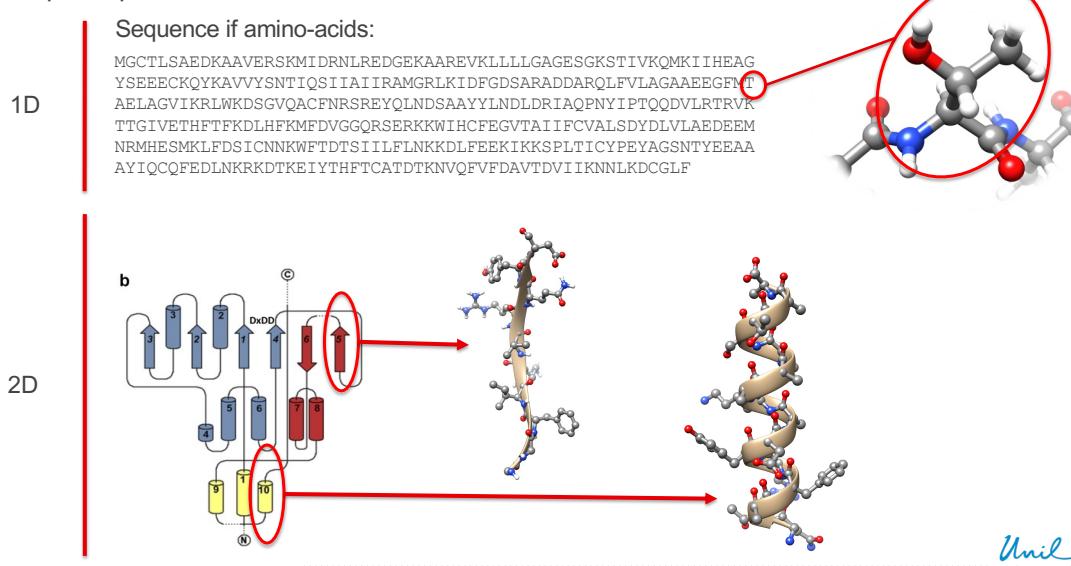
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Molecular representations – biological macromolecules

Biological macromolecules can also be represented in 1D, 2D or 3D:

Example of proteins



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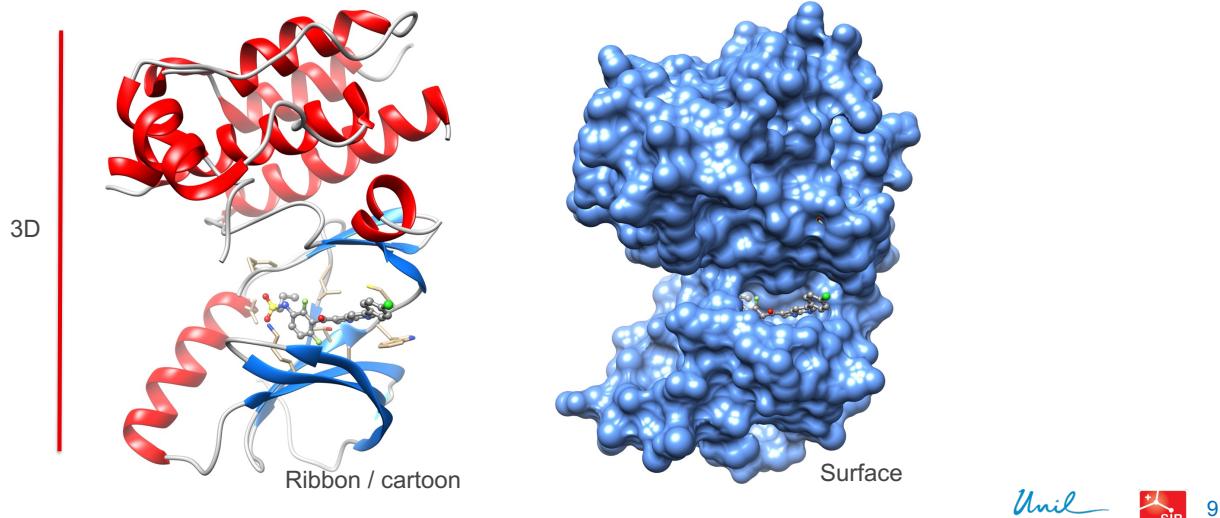
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Molecular representations – biological macromolecules

Biological macromolecules can also be represented in 1D, 2D or 3D:

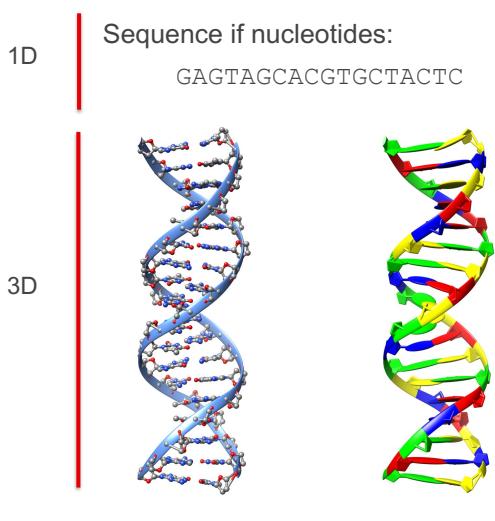
Example of proteins



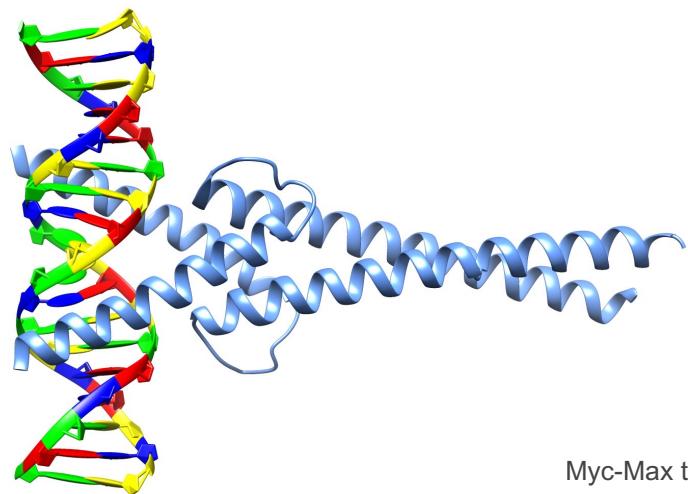
Molecular representations – biological macromolecules

Biological macromolecules can also be represented in 1D, 2D or 3D:

Example of DNA

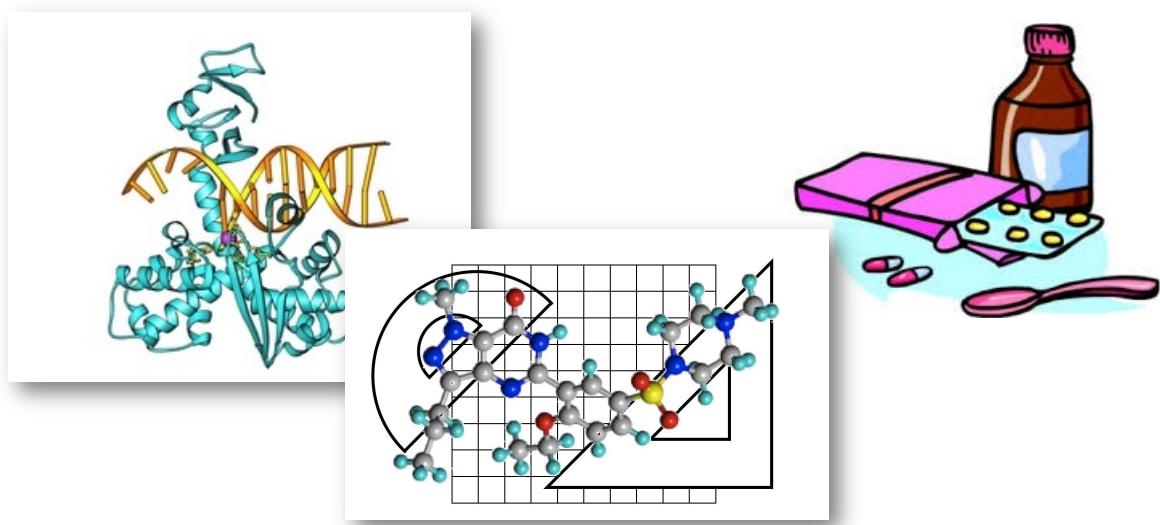


Molecular representations – biological macromolecules



Myc-Max transcription factor

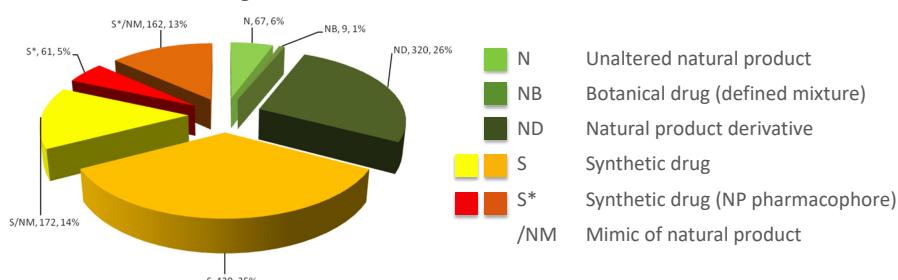
Overview of the Drug Design Pipeline



Drugs: Definition and Origin

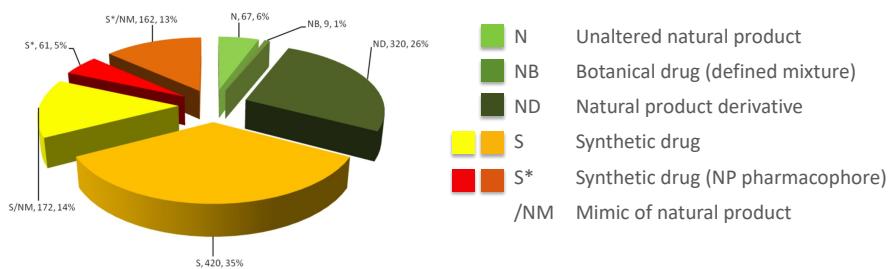
Drug (here = active ingredient):

- A **substance** administered to a patient with possibly various objectives:
 - a **therapeutic** objective (treatment): to cure a **disease**, or
 - a **prophylactic** objective (prevention): to avert the emergence of a **disease**, or
 - a **diagnostic** objective: to identify and monitor a **disease**.
- In the context of **Drug Design**, the substance is a chemical “small” **molecule**.
- Where do these drug molecules come from ?

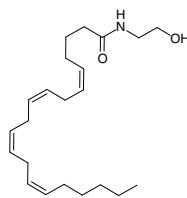


Newman, D. J. & Cragg, G. M. (2016). *Journal of Natural Products*, 79(3), 629.

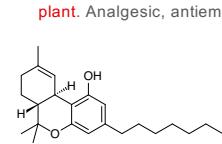
Drugs: Definition and Origin



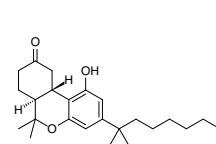
Anandamide. Natural, endogenous, ligand of cannabinoid receptors



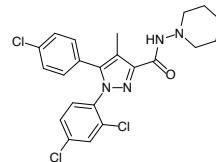
Tetrahydrocannabinol (THC). Natural ligand of cannabinoid receptors, from plant. Analgesic, antiemetic



Nabilone. Synthetic ligand, derived from THC. Analgesic, antiemetic

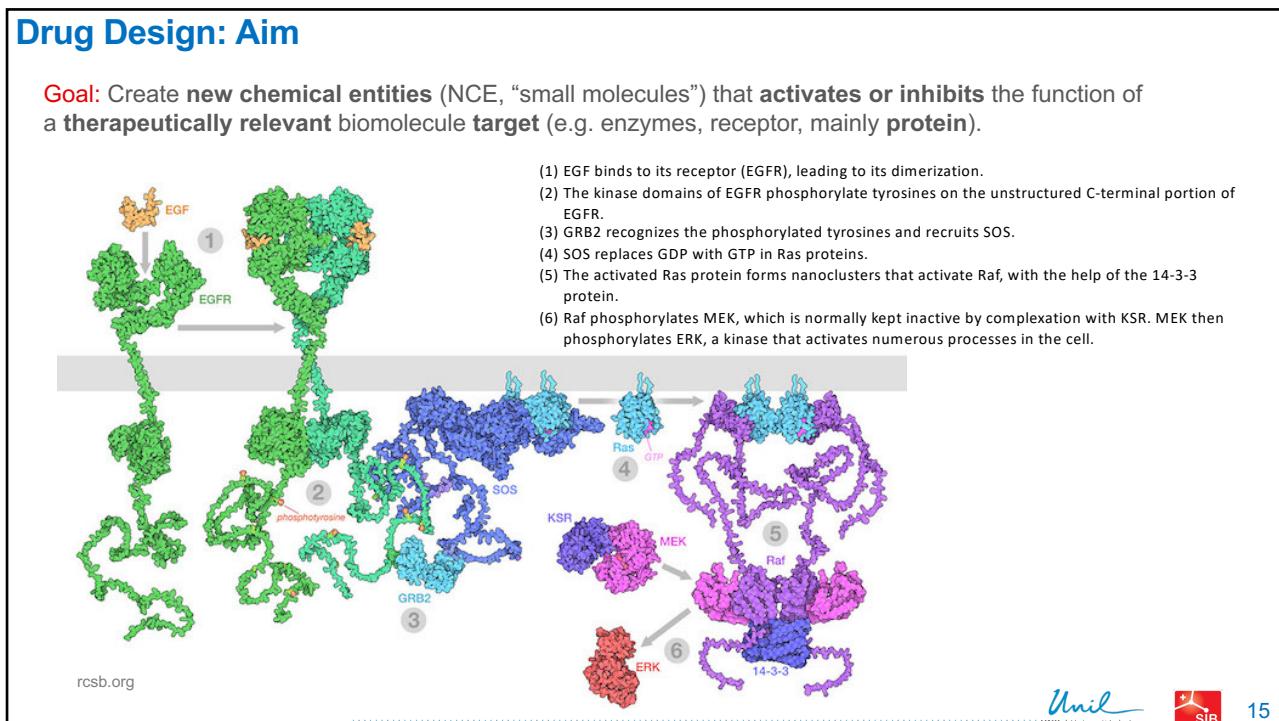


Rimonabant. Synthetic ligand. Anorectic anti-obesity.



Drug Design: Aim

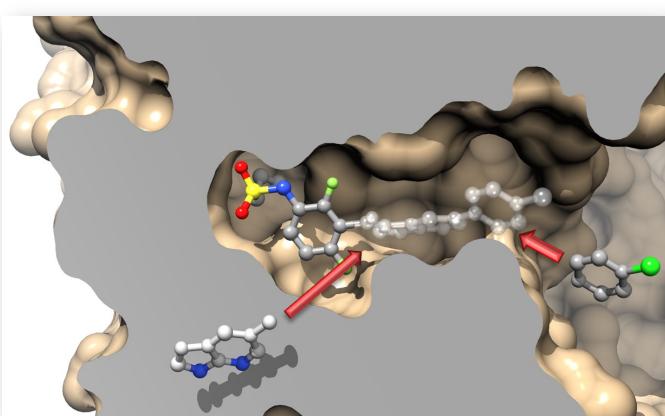
Goal: Create new chemical entities (NCE, “small molecules”) that **activates or inhibits** the function of a therapeutically relevant biomolecule **target** (e.g. enzymes, receptor, mainly protein).



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Drug Design: Aim

Goal: Create new chemical entities (NCE, “small molecules”) that **activates or inhibits** the function of a therapeutically relevant biomolecule **target** (e.g. enzymes, receptor, mainly protein).



To address:

Molecular recognition; i.e.
 “Lock and key” (E. Fischer)

→ Potency, Selectivity

But also **ADMET**,

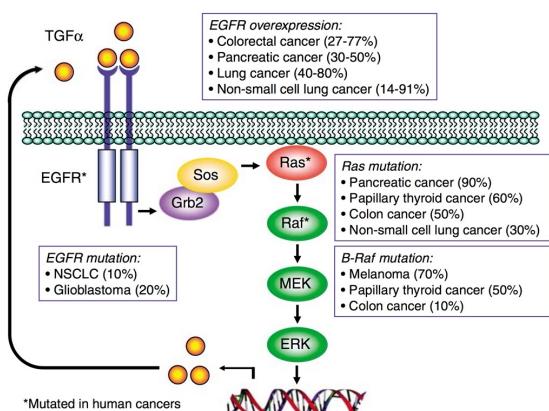
- Absorption
- Distribution
- Metabolism
- Excretion
- Toxicity

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Drug Design: Aim

Goal: Create new chemical entities (NCE, “small molecules”) that **activates or inhibits** the function of a therapeutically relevant biomolecule **target** (e.g. enzymes, receptor, mainly protein).

Possible drugs:



Roberts P.J., et al., *Oncogene* (2007) 26, 3291–3310

EGFR:

Afatinib	Gefitinib	Osimertinib
Almonertinib	Icotinib	Pyrotinib
Brigatinib	Lapatinib	Simotinib
Dacomitinib	Neratinib	Sorafenib
Erlotinib	Olmutinib	Vandetanib

RAS:

Adagrasib	Sotorasib
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RAF:

Dabrafenib	Vemurafenib
Encoratinib	

MEK:

Binimetinib	Selumetinib	Trametinib
Cobimetinib		

ERK:

Ulixertinib	
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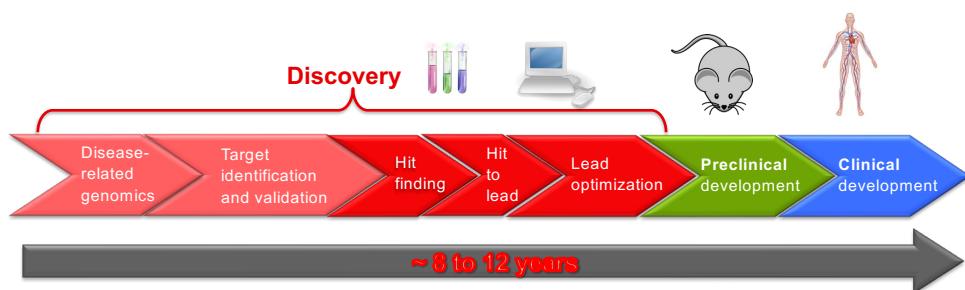


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Drug Design: Pipeline

Goal: Create new chemical entities (NCE, “small molecules”) that **activates or inhibits** the function of a therapeutically relevant biomolecule **target** (e.g. enzymes, receptor, mainly protein).



- **Hit:** molecule showing a signal of activity for the target.
- **Hit finding:** process to discover hits, generally using **Molecular Screening (HTS)**.
- **Hit-to-lead:** Selection of select hits. **Activity confirmation, re-testing** for dose-response. Filters (toxicity, ...).
- **Lead:** molecule showing **promising and confirmed properties**.
- **Lead optimization:** Modest and targeted **chemical modifications** of the lead to **refine** the properties of lead.
- **Preclinical development:** **animal pharmacology/toxicology testing:** reasonably **safe to proceed with human?**
- **Clinical development:** **safety, dosage, efficacy side-effects** in human



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Drug design : some figures

Globally:

- ~ 40 new active ingredients on the market each **year**,
- including 10 'first in class', i.e. drugs with new mode of action.

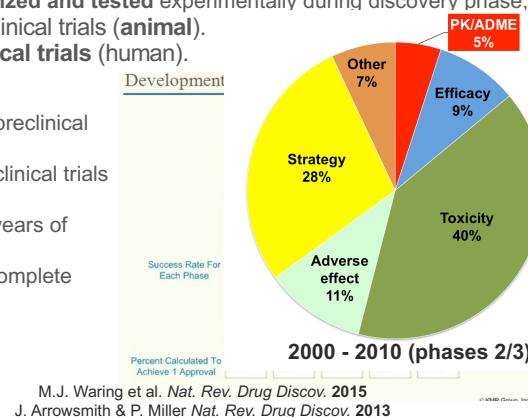
Typical project:

- Millions of chemical structures ("virtual molecules") created and/or evaluated in computer,
- Thousands of molecules synthesized and tested experimentally during discovery phase,
- 3 to 10 molecules tested in preclinical trials (**animal**).
- 1 to 3 molecules to enter in **clinical trials** (**human**).

Outcome, duration and costs:

- 3 to 10% of the molecules entering preclinical trials will become drugs
- 5 to 17% of the molecules entering clinical trials will become drugs
- 8 - 12 years in total, including 6 - 7 years of clinical trials
- Total cost: ~1 billion dollars for a complete project

➔ Risky and expensive.

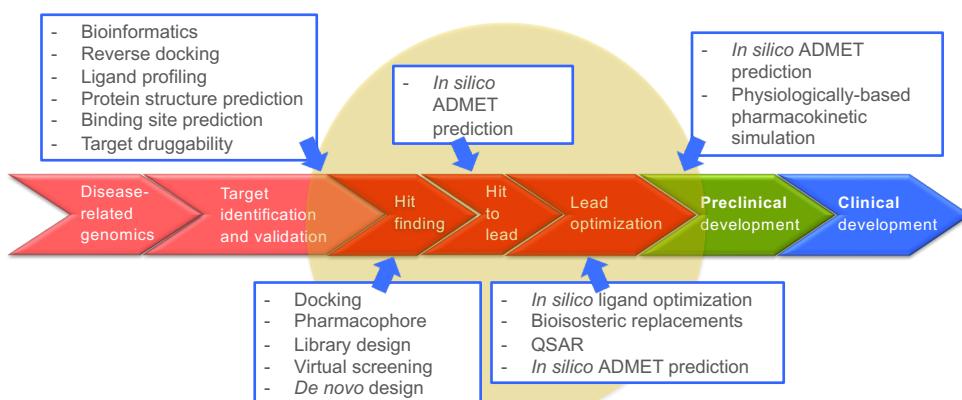


Computer-Aided Drug design (CADD)

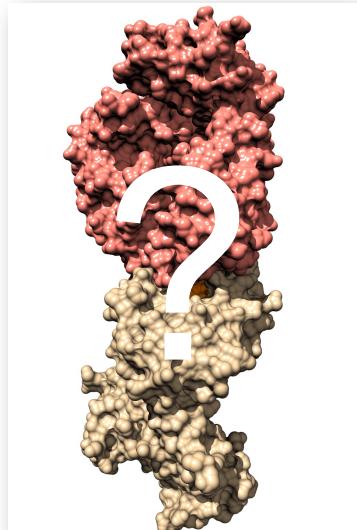
Objective: use of **computing resources**, algorithms and 3D visualization (programs, web-services, databases) to **support**:

- **rational ideas** about how to **create** or **modify** molecules,
- **decisions making** in the execution of the drug design process

CADD is including a lot of different approaches, methods, techniques and tools:



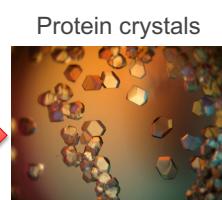
Origin of the 3D structures



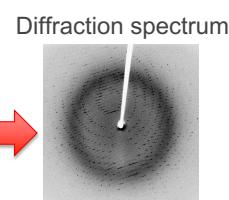
Experimental methods – Xray crystallography



Crystallization

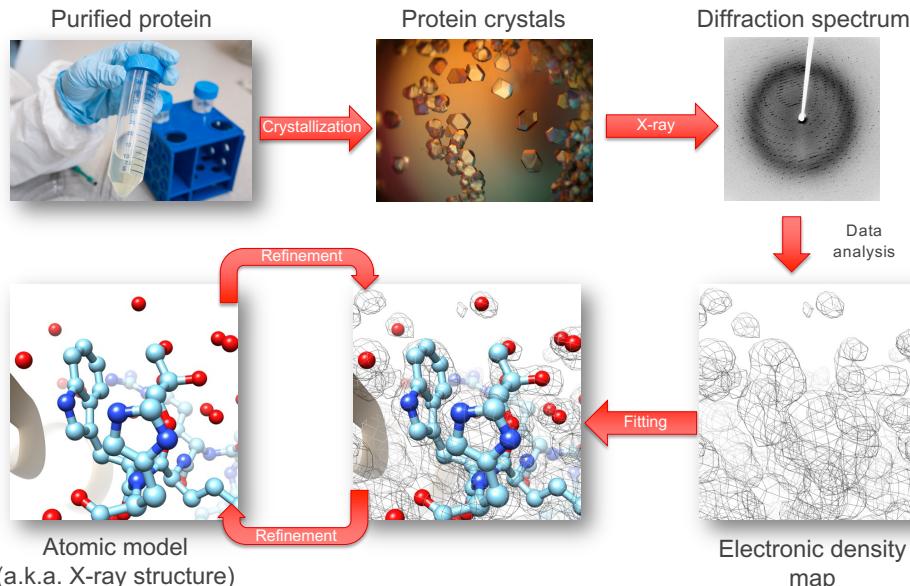


X-ray



Xray diffraction

Experimental methods – Xray crystallography



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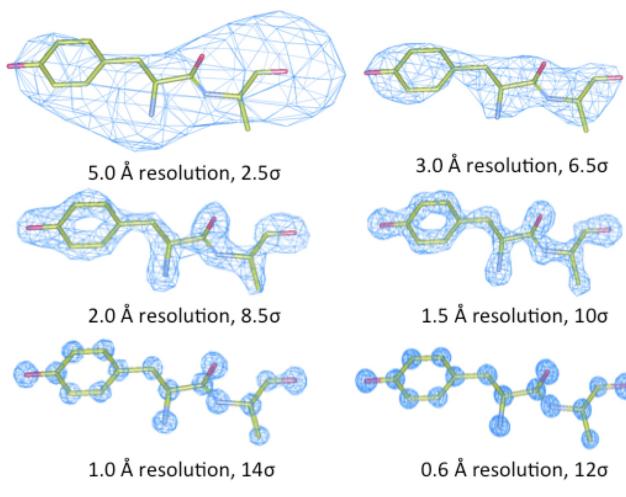
23

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Experimental methods – Xray crystallography

important measures of accuracy:

- **Resolution** (in Å): measures the amount of detail that may be seen in the experimental data. The lower the better (typically around 2 Å)



Source: PLoS One. 2015 Apr 20;10(4):e0123146.

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Experimental methods – Xray crystallography

3 important measures of accuracy:

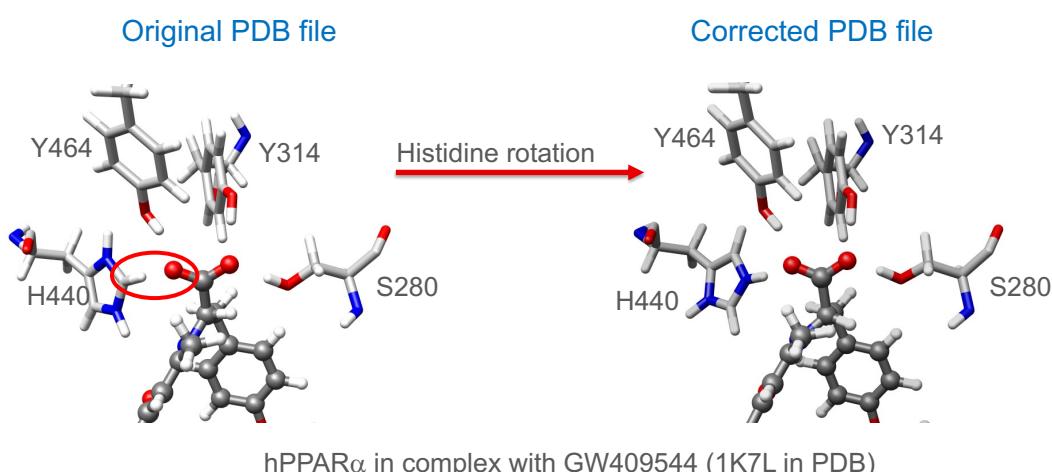
- **Resolution** (in Å): measures the amount of detail that may be seen in the experimental data. The lower the better (typically around 2 Å)
- **R-value**: measures how well the atomic model is supported by the experimental data found in the structure factor file (Perfect fit R-value = 0.0; Random fit R-value = 0.63; Typical R-value ~ 0.20) The atomic model is used to simulate a diffraction spectrum, which is compared to the experimental one.
- **R-free value**: idem than R-value, but calculated for a set of experimental data that have not been used to create the model (~10% of the data are removed before refinement, in order to be used in this test). Generally, R-free value > R-value; Typically R-free value ~ 0.26 for a good quality structure.

Typical limitations:

- Hydrogen atoms are generally not visible
- Some regions are not defined (e.g. flexible loops or flexible side chains)
- X-ray structures are models. They can be totally wrong!!

Experimental methods – Xray crystallography

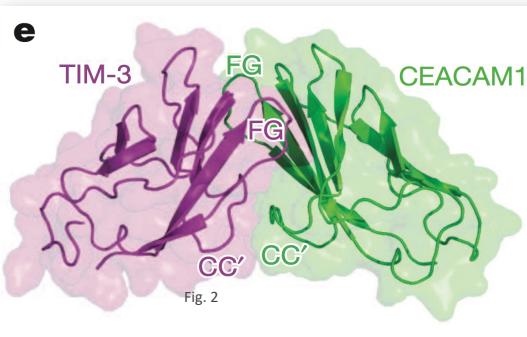
Xray structures **are models**. They can be wrong!



Experimental methods – Xray crystallography

Xray structures **are models**. They can be totally wrong!

Huang, Y.-H., et al. CEACAM1 regulates TIM-3-mediated tolerance and exhaustion. *Nature*, 2015, 517(7534), 386–390.



Xray structure of the complex
CEACAM1/TIM3
PDB ID: 4QYC
Resolution: 3.4Å
R-value: 0.232

Correction →

5DZL

Crystal structure of the protein human CEACAM1

DOI: 10.2210/pdb5dzl/pdb Entry 5DZL supersedes 4QYC

It was a homodimer of CEACAM1...!

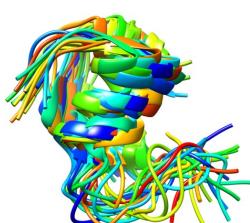
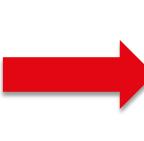
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Experimental methods – NMR spectroscopy

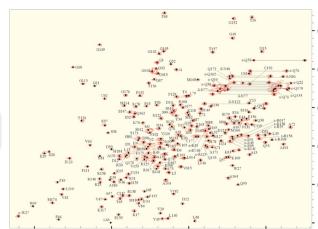


Purification
Concentration



Ensemble of structures
Example: insulin

Distance
constraints
Modeling



2D-Spectra

Pros : Structure in solution
Cons: - Limited to small proteins
- Low resolution
- Highly flexible regions

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Experimental methods – CryoEM

DUBOCHET'S VITRIFICATION METHOD

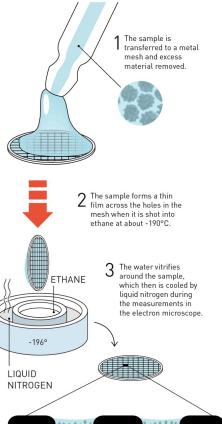
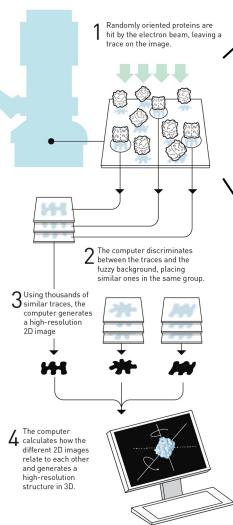


Illustration: ©Johan Jarnestad/The Royal Swedish Academy of Sciences

FRANK'S IMAGE ANALYSIS FOR 3D STRUCTURES



- Very power electronic beam
- Better resolution than light (smaller wave length)
- In vacuo in the microscope
- Frozen sample (77 K or 4 K)
- Vitrified water



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Experimental methods – CryoEM

DUBOCHET'S VITRIFICATION METHOD

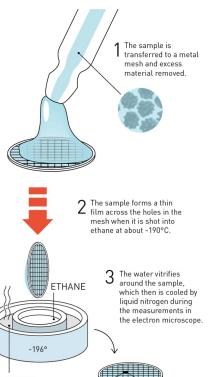
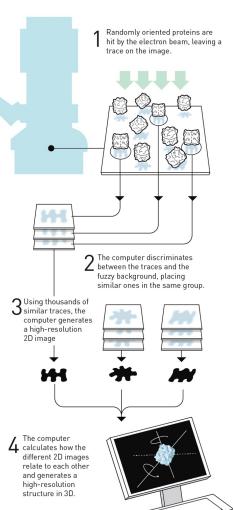


Illustration: ©Johan Jarnestad/The Royal Swedish Academy of Sciences

FRANK'S IMAGE ANALYSIS FOR 3D STRUCTURES



Until recently:

- Only low resolution structures. Need to be used together with Xray crystallography or NMR (for example, insertion of Xray structures into the Cryo-EM density map)
- Limited to large-size systems (which can actually be seen as a pros or a cons)

Nowadays:

- Resolution close to that of Xray crystallography
- Applicable to smaller systems
- More Cryo-EM structures produced every year than NMR structures
- Capture structures in relevant states (isolated molecules, in solution, at a given salt concentration and pH)

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Experimental methods - Summary

Technique	Advantages	Disadvantages
Xray crystallography	High resolution (1 to 3 Å)	Requires to crystallize the protein Does not allow studying transmembrane or very flexible proteins
NMR	Does not require protein crystallization ~ High resolution	Generally limited to small proteins
Cryo-EM	Does not necessitate to crystallize the protein: possible to study transmembrane proteins, and more flexible proteins than Xray. New techniques allow studying smaller proteins, and increasing resolution	Generally limited to large proteins Low resolution, 4 to 20 Å (a lot of progresses have been done recently)

Where to find experimental 3D structures? The protein databank

Public experimental 3D structures are stored in the **Protein Data Bank (PDB)**

Worldwide Protein Data Bank (wwPDB)

RCSB Protein Data Bank (RCSB PDB)
Protein Data Bank in Europe (PDBe)
Protein Data Bank Japan (PDBj)

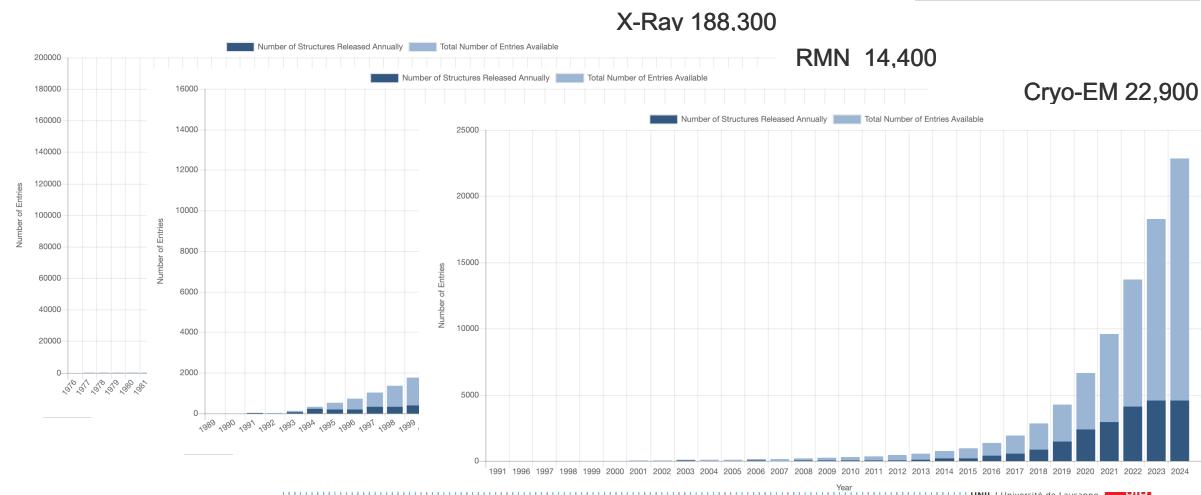
<https://www.wwpdb.org>

<https://www.rcsb.org>

<https://www.ebi.ac.uk/pdbe>

<https://pdbj.org>

226'000 structures
in Oct 2024



Where to find experimental 3D structures? The protein databank

<https://www.rcsb.org>

The screenshot shows the RCSB PDB homepage. At the top, a search bar is highlighted with the text "C-MET crizotinib". Below the search bar, the RCSB PDB logo and statistics are displayed: 225,946 Structures from the PDB and 1,068,577 Computed Structure Models (CSM). The main content area features a "Welcome" sidebar with links to Deposit, Search, Visualize, Analyze, Download, and Learn. A central banner highlights "Experimentally-determined 3D structures from the Protein Data Bank (PDB) archive" and "Computed Structure Models (CSM) from AlphaFold DB and ModelArchive". A "Explore NEW Features" section shows a computer monitor with a "NEW!" button and a lightbulb icon. To the right, a "October Molecule of the Month" section shows a 3D ribbon model of a protein complex, identified as "Angiotensin and Blood Pressure". The bottom right corner features logos for UNIL, SIB, and 33.

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Where to find experimental 3D structures? The protein databank

The screenshot shows the search results page of the RCSB PDB. On the left, a sidebar contains filters for PDB ID, Authors, Experimental methods, Refinement Resolution (Å), and Release Date. The main area displays three search results: 2XP2 (Structure of the Human Anaplastic Lymphoma Kinase in Complex with Crizotinib), 2WGJ (X-ray Structure of PF-02341066 bound to the kinase domain of c-Met), and 3ZBF (Structure of Human ROS1 Kinase Domain in Complex with Crizotinib). Each result includes a 3D ribbon model, a "Explore in 3D" button, and detailed information about the structure, including PDB ID, authors, release date, method, organisms, macromolecule, and unique ligands. A red box highlights the "Sort by" dropdown menu in the top right of the results table, with the text "Possible to sort" written next to it. The bottom right corner features logos for UNIL, SIB, and 34.

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Where to find experimental 3D structures? The protein databank

Refinements

Structure Determination Methodology

experimental (157)

Scientific Name of Source Organism

Homo sapiens (151)

Listeria monocytogenes EGD-e (8)

synthetic construct (3)

Gallus gallus (1)

Mus musculus (1)

Taxonomy

Eukaryota (152)

Bacteria (8)

other sequences (3)

Eukaryotes (eukaryotes) (1)

Experimental Method

X-RAY DIFFRACTION (51)

ELECTRON MICROSCOPY (5)

SOLUTION NMR (1)

Polymer Entity Type

Protein (157)

Refinement Resolution (Å)

1.0 - 1.5 (6)

1.5 - 2.0 (63)

2.0 - 2.5 (59)

2.5 - 3.0 (19)

3.0 - 3.5 (4)

4.0 - 4.5 (2)

> 4.5 (9)

Release Date

1995 - 1999 (1)

2000 - 2004 (4)

2005 - 2009 (25)

2010 - 2014 (57)

1 to 25 of 157 Structures

Page 1 of 7 | 25 | Sort by Score

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2XP2
Structure of the Human Anaplastic Lymphoma Kinase in Complex with Crizotinib (PF-02341066)
McTigue, M., Deng, Y., Liu, W., Broun, A., Timofeevski, S., Marone, T., Cui, J.J.
(2011) J Med Chem 54: 6342

Released 2010-09-15
Method X-RAY DIFFRACTION 1.9 Å
Organisms Homo sapiens
Macromolecule TYROSINE-PROTEIN KINASE RECEPTOR (protein)
Unique Ligands VGH

2WGJ
X-ray Structure of PF-02341066 bound to the kinase domain of c-Met
McTigue, M., Grodsky, N., Ryan, K., Tran-Dube, M., Cui, J.J., Mroczkowski, B.
(2011) J Med Chem 54: 6342

Released 2009-06-02
Method X-RAY DIFFRACTION 2 Å
Organisms Homo sapiens
Macromolecule HEPATOCYTE GROWTH FACTOR RECEPTOR (protein)
Unique Ligands VGH

3ZBF
Structure of Human ROS1 Kinase Domain in Complex with Crizotinib
McTigue, M., Deng, Y., Liu, W., Broun, A., Stewart, A.
(2013) N Engl J Med 368: 2395

Released 2013-06-12
Method X-RAY DIFFRACTION 2.2 Å
Organisms Homo sapiens
Macromolecule PROTO-ONCOGENE TYROSINE-PROTEIN KINASE ROS (protein)
Unique Ligands VGH

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Where to find experimental 3D structures? The protein databank

Structure Summary | Structure | Annotations | Experiment | Sequence | Genome | Ligands | Versions

2WGJ
X-ray Structure of PF-02341066 bound to the kinase domain of c-Met
PDB DOI: <https://doi.org/10.2210/pdb2WGJ/pdb>

Classification: TRANSFERASE
Organism(s): Homo sapiens
Expression System: Spodoptera frugiperda
Mutation(s): No

Released: 2009-04-20 | Deposited: 2009-06-02
Deposited Author(s): McTigue, M., Grodsky, N., Ryan, K., Tran-Dube, M., Cui, J.J., Mroczkowski, B.

Experimental Data Snapshot
Method: X-RAY DIFFRACTION
Resolution: 2.0 Å
R-Value Free: 0.232
R-Value Work: 0.214
R-Value Observed: 0.215

Starting Model: experimental
View more details

wwPDB Validation
Method: X-RAY DIFFRACTION
Resolution: 2.0 Å
R-Value Free: 0.232
R-Value Work: 0.214
R-Value Observed: 0.215

Ligand Structure Quality Assessment
Worse | Better

Download Primary Citation | Download File | Data API

Organism (origin of the sequence) & expression system (synthesis et post-translational modifications)

Note: post-translational modifications can differ between organisms

Dowload or online visualization

Experimental method and quality

Structure Based Drug Design of Crizotinib (PF-02341066), a Potent and Selective Dual Inhibitor of Mesenchymal-Epithelial Transition Factor (c-Met) Kinase and Anaplastic Lymphoma Kinase (ALK).
Cui, J.J., Tran-Dube, M., Shen, H., Nambu, M., Kung, P.P., Parish, M., Jia, L., Meng, J., Funk, L., Botros, I., Mroczkowski, M., Grodsky, N., Ryan, K., Padraig, E., Alton, G., Timofeevski, S., Yamazaki, S., Li, Q., Zou, H., Cahnman, L., Mroczkowski, B., Bender, S., Kania, R.S., Edwards, M.P.
(2011) J Med Chem 54: 6342

PubMed ID: 21812414 | DOI: <https://doi.org/10.1021/j102007613>
Primary Citation of Related Structures:
2WGJ, 2WKM, 2WQJ

PubMed Abstract:
Because of the critical roles of aberrant signaling in cancer, both c-MET and ALK receptor tyrosine kinases are attractive oncology targets for therapeutic intervention. The co crystal structure of 3 (PfHA-665752), bound to c-MET kinase domain, revealed a novel ATP site environment, which served as the target to guide parallel...

Download Primary Citation | Download File | Data API

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Online visualization

2WGJ.

X-ray Structure of PF-02341066 bound to the kinase domain of c-Met

37

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Information regarding the protein, and what is present in the experimental structure

Macromolecules

Find similar proteins by: [Sequence](#) (by identity cutoff) | [3D Structure](#)

Entity ID: 1

Molecule	Chains	Sequence Length	Organism	Details	Image
HEPATOCYTE GROWTH FACTOR RECEPTOR	A	306	Homo sapiens	Mutation(s): 0 (0) EC: 2.7.10.1	

UniProt & NIH Common Fund Data Resources

Find proteins for [P08581](#) (Homo sapiens) Explore [P08581](#) (0) Go to UniProtKB [P08581](#)

PHAROS: [P08581](#) PTEX: [ENSG00000105976](#)

Entity Groups: [Sequence Clusters](#) (30% Identity, 40% Identity, 70% Identity, 90% Identity, 95% Identity, 100% Identity) [UniProt Group](#) [P08581](#)

Sequence Annotations

Reference Sequence: 2WGJ_1

2WGJ_1
UNIPROT P08581
UNMOEDED

HYDROPHITY

DISORDER

DISORDERED BINDING

PFAM

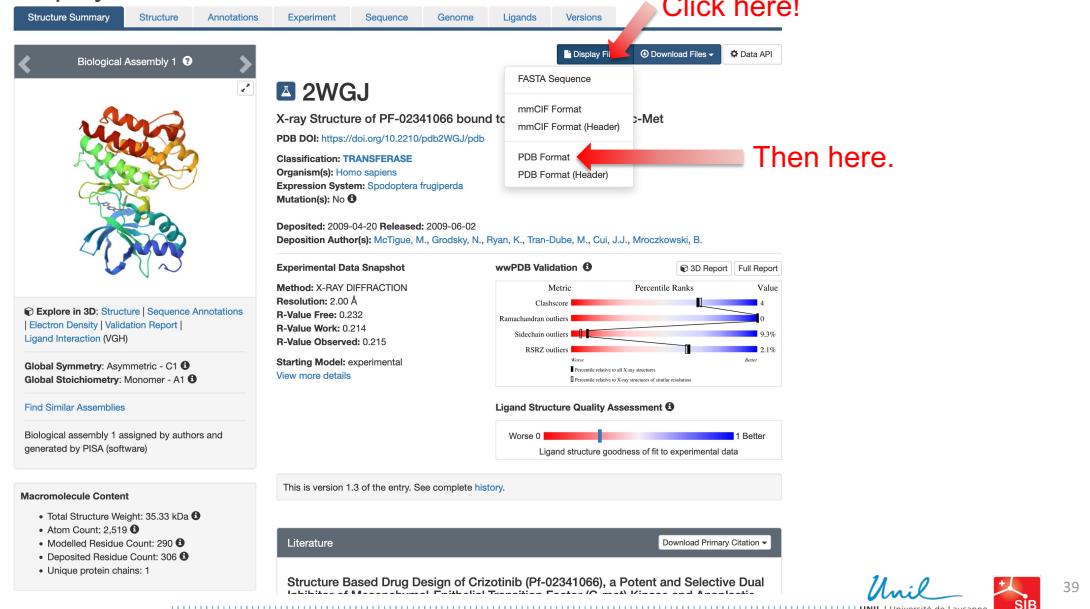


Expand

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Where to find experimental 3D structures? The protein databank

Download / display



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Header with information about the protein and experimental conditions

20-APR-09 2W6J
 X-RAY STRUCTURE OF PF-02341666 BOUND TO THE KINASE DOMAIN OF C-MET
 CPNDID: 1
 2 MOLECULE: HEPATOCYTE GROWTH FACTOR RECEPTOR;
 3 HGR; HGF; MET; METABotropic GLUTAMATE RECEPTOR 1; MGR; MET-1;
 4 FRAGMENT: TYROSINE KINASE DOMAIN, RESIDUES 181-1348;
 5 SYNTHON: HGF RECEPTOR, SCATTER FACTOR RECEPTOR, SF RECEPTOR, HGF/SF
 6 PROTEIN: PROTEIN-OXYGENIC TYROSINE KINASE, C-MET;
 7 ECT: 2.7-18.1 Å
 8 ENGINEERED: YES
 9 SOURCE: 1 ORGANISM: SCIENTIFIC; HOMO SAPINENSIS;
 10 SOURCE: 2 ORGANISM: COMMON; HUMAN;
 11 SOURCE: 3 ORGANISM: COMMON; HUMAN;
 12 SOURCE: 4 ORGANISM: COMMON; HUMAN;
 13 SOURCE: 5 EXPRESSION SYSTEM: SP2ODERMA FRUGIPERDA;
 14 SOURCE: 6 EXPRESSION SYSTEM: TETRASOMA TETRASOMA;
 15 SOURCE: 7 EXPRESSION SYSTEM: TETRASOMA TETRASOMA;
 16 SOURCE: 8 EXPRESSION SYSTEM, VECTOR TYPE: BACULIVIRUS;
 17 SOURCE: 9 EXPRESSION SYSTEM, PLASMID: PFBTAC1;
 18 KEYWORD: 1 TYROSINE KINASE; 2 PROTEIN-OXYGENIC, ATP-BINDING, NUCLEOTIDE-
 19 KEYWORD: 2 BINDING, TYROSINE-PROTEIN KINASE
 20 AX-RAY DIFFRACTION
 21 EXP: 1 15-DEC-23 2W6J 1 REMARK
 22 EXP: 2 08-NOV-19 2W6J 1 REMARK
 23 EXP: 3 20-SEP-11 2W6J 1 REMARK JNLN. REMARK FORMUL
 24 EXP: 4 01-SEP-10 2W6J 1 VERSN
 25 EXP: 5 01-SEP-10 2W6J 1 REMARK
 26 EXP: 6 01-SEP-10 2W6J 1 REMARK
 27 EXP: 7 02-JUN-09 2W6J 1 REMARK
 28 JNLN: AUT: A. BURSHUDOV, M. H. BIEB, P. KANG, M. PARHESH, L. JIA,
 29 JNLN: AUT: 1 ZHANG, L. FUNK, I. BUDROV, M. MCTIGUE, N. GROSKY, O. LI, H. ZOU
 30 JNLN: AUT: 2 E. CHRISTENSEN, B. W. ZIMKINS, J. S. R. S. K. M. PARHESH, P. KANG,
 31 JNLN: TITL: 1 THE TYROSINE KINASE INHIBITOR PF-02341666 (PF-02341666) AS A
 32 JNLN: TITL: 2 A POTENTIAL AND SELECTIVE DUAL INHIBITOR OF
 33 JNLN: TITL: 3 A NEUTROPHILIC-TRANSFORMING FACTOR (C-MET) KINASE AND
 34 JNLN: TITL: 4 AN ELLIPTICATIC LYMPHOMA KINASE (ALK)
 35 JNLN: REF: J.MED.CHEM 54 6342 2011
 36 JNLN: PDB: 2W6J
 37 JNLN: PDBID: 2W6J
 38 JNLN: ISSN: 0882-6263
 39 JNLN: PROTEIN: 2B12A1
 40 JNLN: DOI: 10.1016/j.jmedchem.2011.03.013
 41 REMARK 2 2.00 ÅNGSTROMS
 42 REMARK 3
 43 REMARK 3 REFINEMENT: P = REFMAC 5.1.24
 44 REMARK 3
 45 REMARK 3 AUTHORS: MURSHUDOV, SHUBAK, LEBEDEV, PANNU, STEINER,
 46 REMARK 3 NICHOLLS, LONG, VASIN
 47 REMARK 3 REFINEMENT TARGET : MAXIMUM LIKELIHOOD
 48 REMARK 3
 49 REMARK 3 DATA USED IN REFINEMENT:
 50 REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS): 2.40
 51 REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS): 19.96
 52 REMARK 3 DATA CUTOFFS (Å): 3.0
 53 REMARK 3 STO(M(F)): NULL
 54 REMARK 3 COMPLETENESS FOR RANGE (%) : 78.1
 55 REMARK 3 NUMBER OF REFLECTIONS : 17195
 56 REMARK 3

Cartesian coordinates for each visible atom

Atom
number

Atom

Residue

Pept.
Chain
code

Residue
number

B-factor.
Measures the variability of the atom's position.
The higher the value, the more fluctuating the position of this atom

Need visualization software

Ex.: Swiss PDB Viewer, UCSF ChimeraX, Pymol

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40

20

Where to find experimental 3D structures? The protein databank

And for small molecules?

Where to find experimental 3D structures? The protein databank

And for small molecules?

Small Molecules

Ligands 1 Unique				
ID	Chains	Name / Formula / InChI Key	2D Diagram	3D Interactions
VGH Query on VGH	B [auth A]	3-[(1R)-1-(2,6-dichloro-3-fluorophenyl)ethoxy]-5-(1-piperidin-4-yl-1H-pyrazol-4-yl)pyridin-2-amine C ₂₁ H ₂₂ Cl ₂ FN ₅ O KTEIFNKAUNYNNU-GFCCVEGCSA-N		Interactions ▾ Interactions & Density ▾
Binding Affinity Annotations				
ID	Source	Binding Affinity		
VGH	BindingDB: 2WGJ	Ki: min: 2, max: 19 (nM) from 3 assay(s) Kd: min: 0.2, max: 2.1 (nM) from 5 assay(s) IC50: min: 0.51, max: 20 (nM) from 24 assay(s)		
	PDBBind: 2WGJ	Ki: 2 (nM) from 1 assay(s)		

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41

41

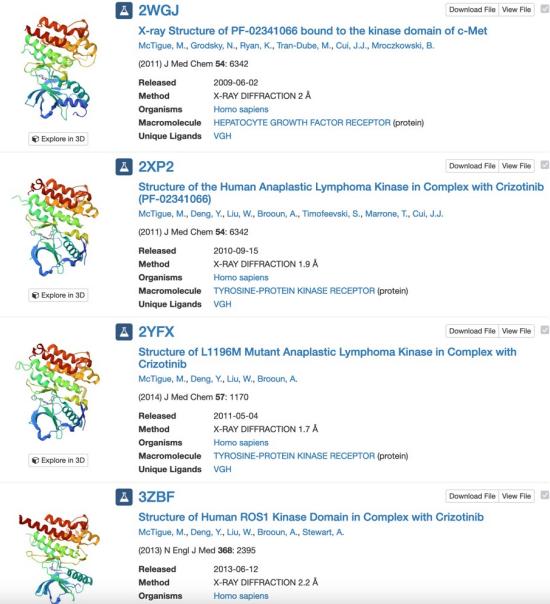
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Where to find experimental 3D structures? The protein databank

List of 3D structures, present in the PDB, and containing the ligand crizotinib



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Where to find experimental 3D structures? The protein databank

Small Molecules

Ligands 1 Unique				
ID	Chains	Name / Formula / InChI Key	2D Diagram	3D Interactions
VGH Query on VGH	B [auth A]	3-[(1R)-1-(2,6-dichloro-3-fluorophenyl)ethoxy]-5-(1-piperidin-4-yl-1H-pyrazol-4-yl)pyridin-2-amine C ₂₁ H ₂₂ Cl ₂ F N ₅ O KTEIFNKAUNYNJU-GFCCVEGCSA-N		<input type="checkbox"/> Interactions <input type="checkbox"/> Interactions & Density
Click here!				

Binding Affinity Annotations		
ID	Source	Binding Affinity
VGH	BindingDB: 2WGJ	K _i : min: 2, max: 19 (nM) from 3 assay(s) K _d : min: 0.2, max: 2.1 (nM) from 5 assay(s) IC ₅₀ : min: 0.51, max: 20 (nM) from 24 assay(s)
	PDBBind: 2WGJ	K _i : 2 (nM) from 1 assay(s)

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44

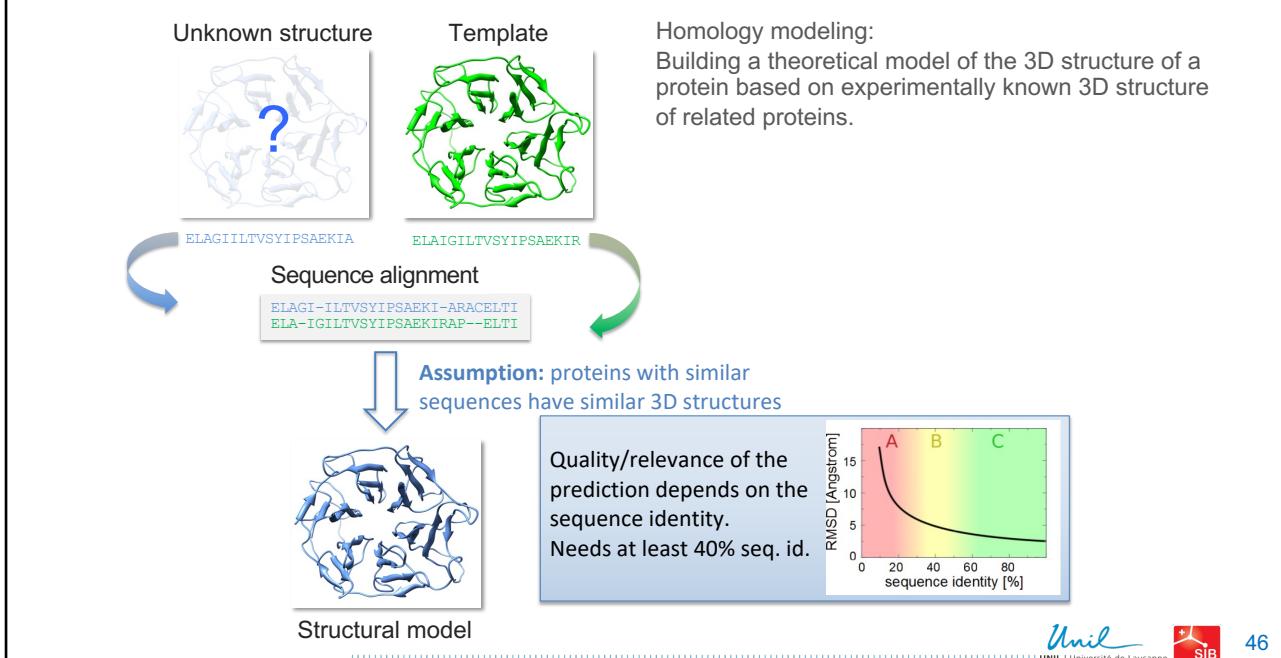
Where to find experimental 3D structures? The protein databank



45

45

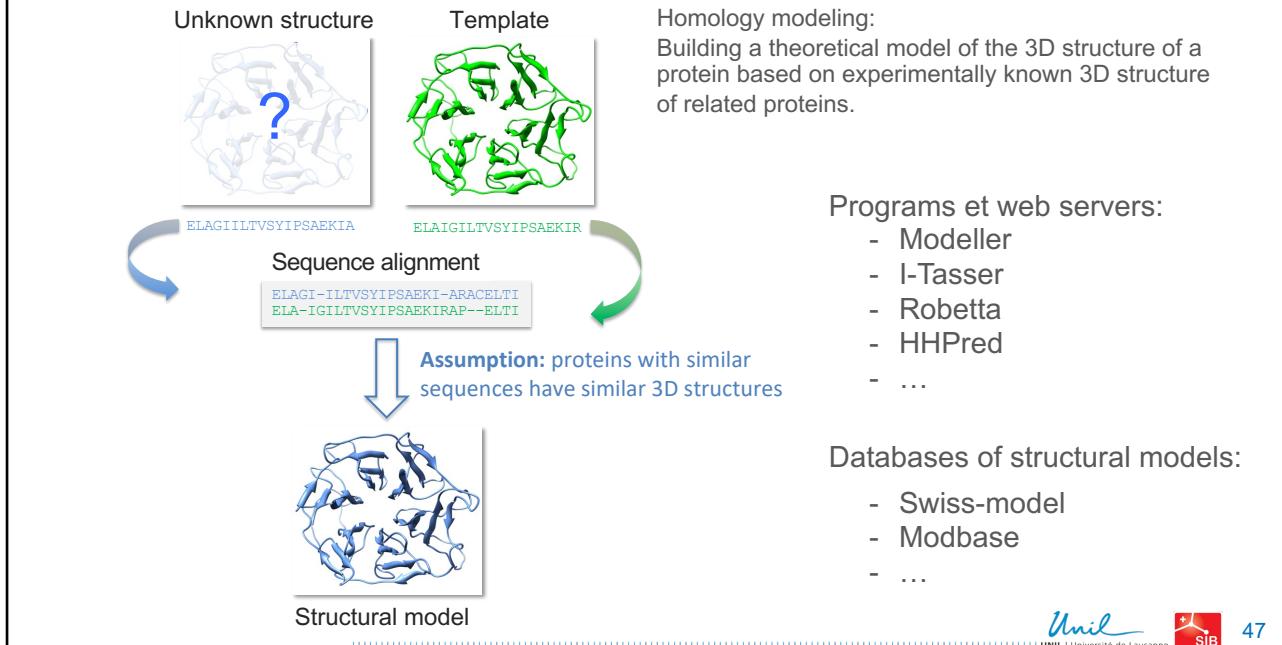
And when there is no experimental structure? Homology modeling



46

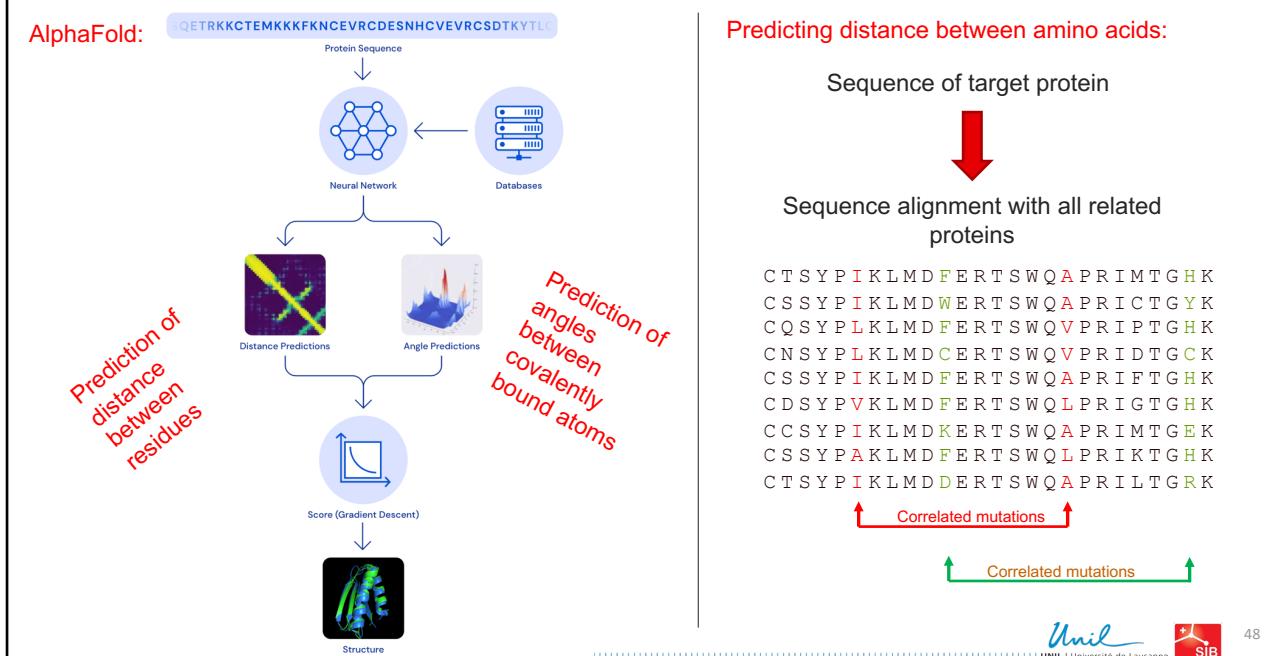
46

And when there is no experimental structure? Homology modeling



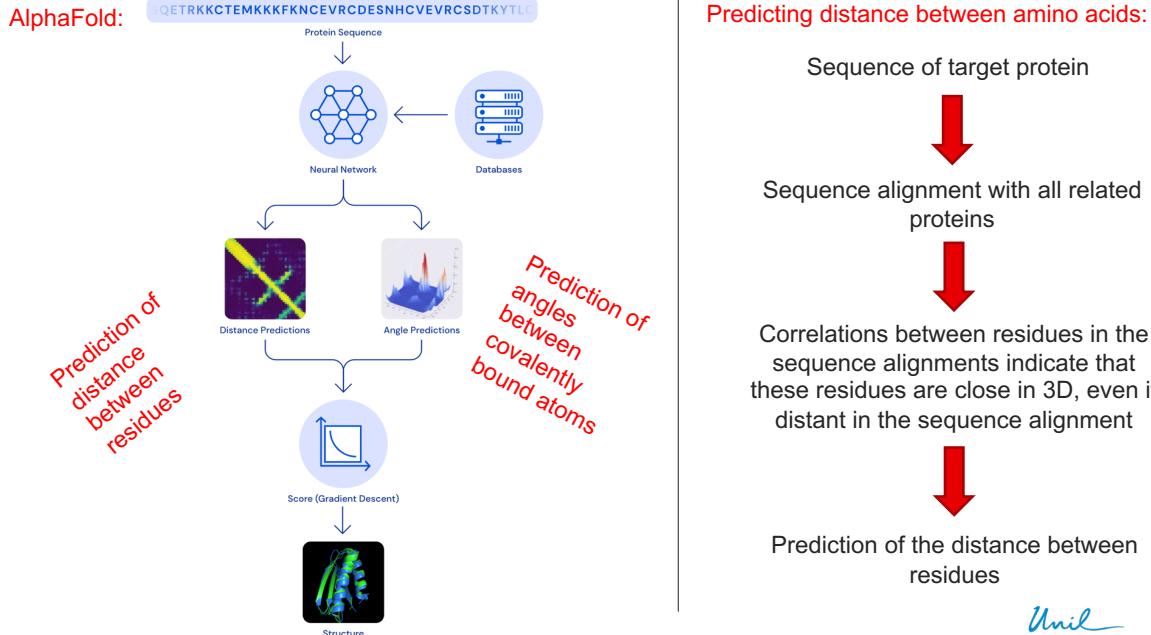
47

And when there is no experimental structure? Homology modeling



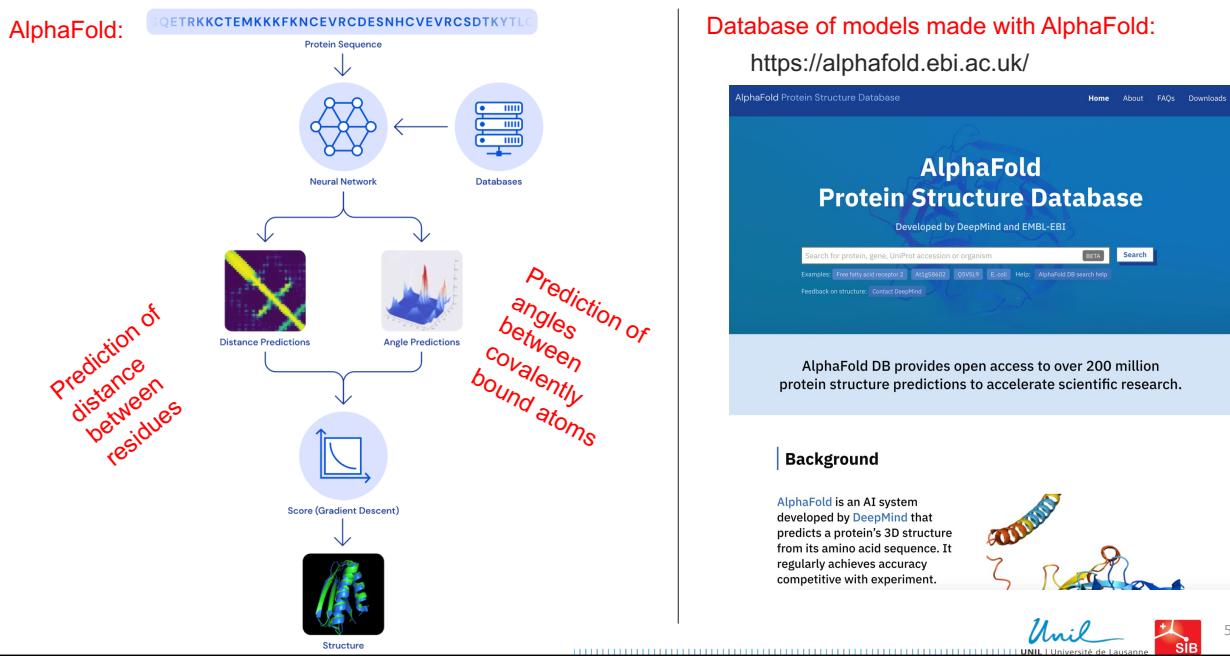
48

And when there is no experimental structure? Homology modeling



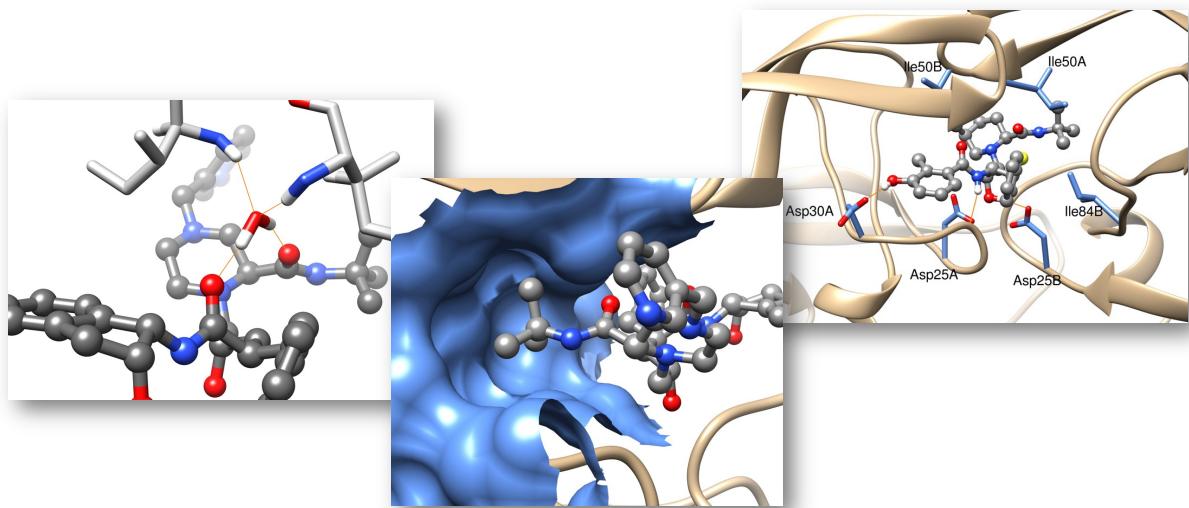
49

And when there is no experimental structure? Homology modeling



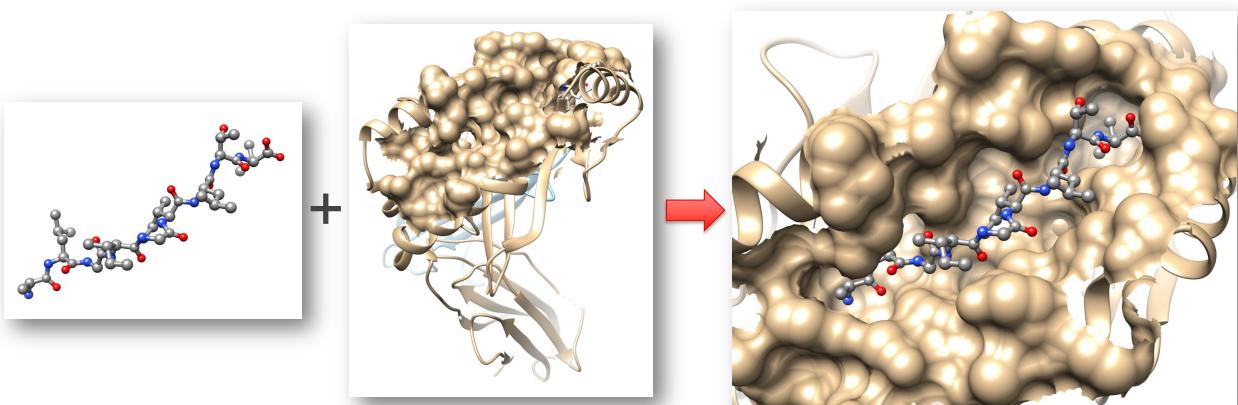
50

Molecular Recognition



Molecular recognition

Molecular interactions → Molecular recognition → Biological response



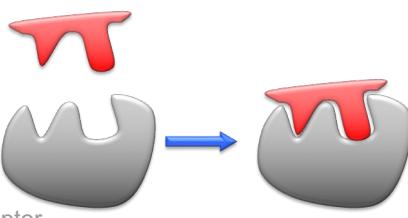
Molecular recognition – Historical models

“Lock and key” model.

Emil Fischer in the 1890s.

The protein has a particular shape into which the ligand fits exactly.

Ligand

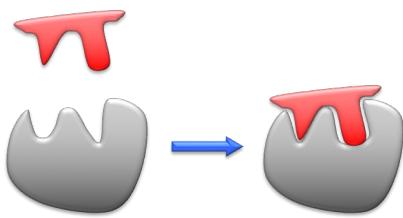


Receptor

Induced fit model

Daniel Koshland 1958.

The binding site of the macromolecule is flexible and its shape can be modified as the ligand interacts with it.



Molecular recognition:

Collection of **interactions** between molecules that govern their **binding**.

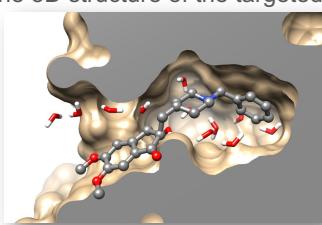
Qualitative **nature** of the interactions?

Quantitative **intensity** of the molecular recognition?

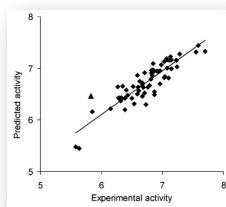
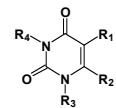
Molecular recognition and CADD

Two main categories of CADD approaches to discover, create, optimize and evaluate active molecules:

- **Structure-based approaches.** Use the 3D structure of the targeted macromolecule. Ex: Molecular docking.



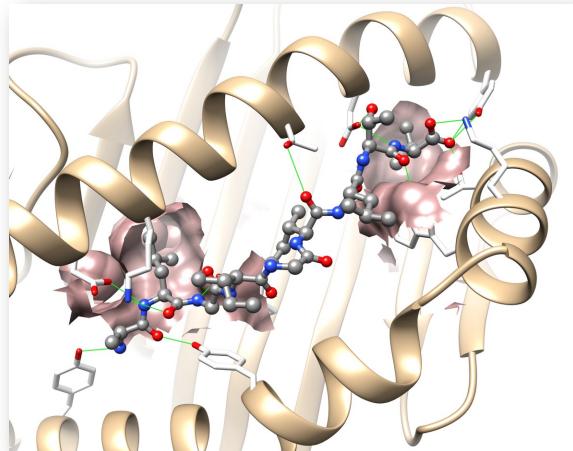
- **Ligand-based approaches.** Use the information derived from known ligands. Ex: Quantitative Structure-Activity Relationships (QSAR), bioisosteric replacements.



Molecular recognition - type of interactions

Non covalent interactions between atoms :

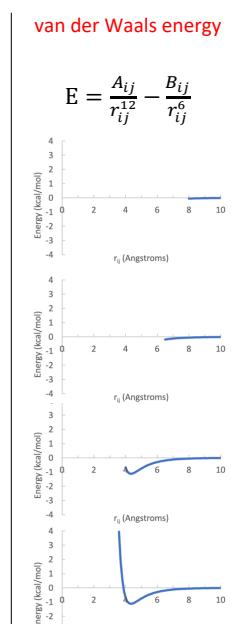
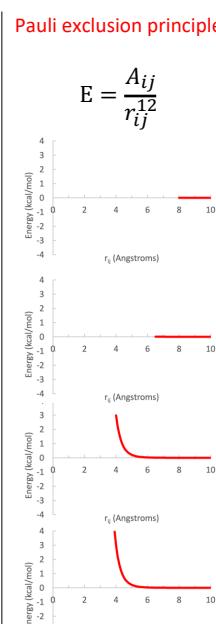
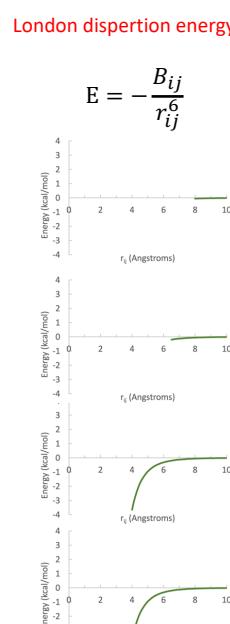
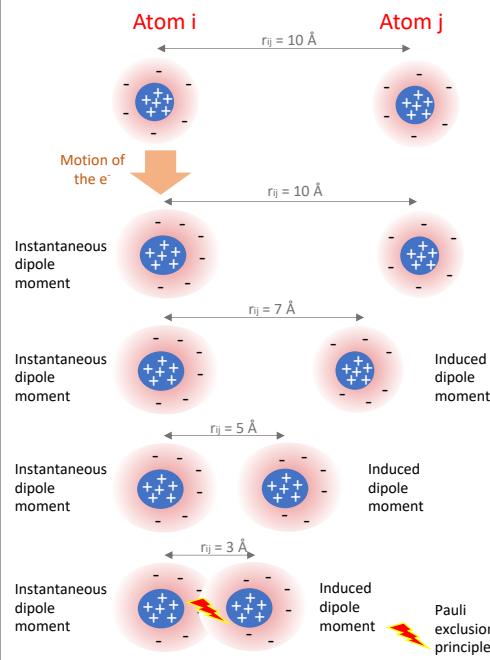
- non-polar interactions (shape recognition)
- electrostatic interactions (salt bridge and hydrogen bond)
- π interactions
- metal/ion interactions



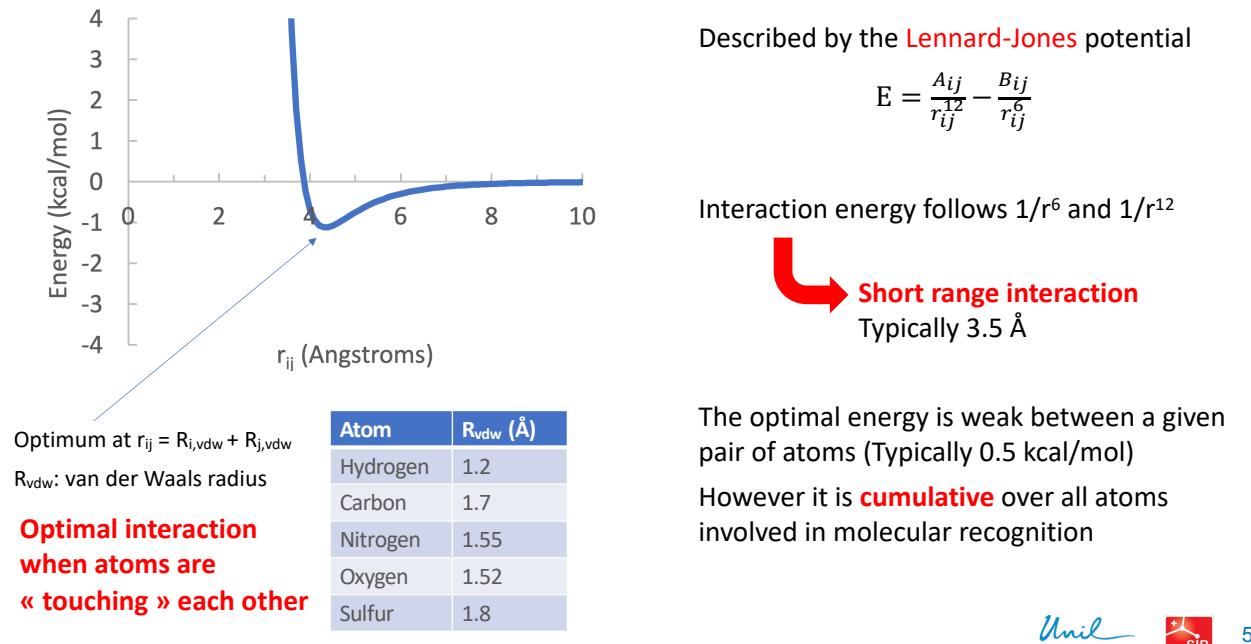
Crystal structure of HLA-A2*0201 in complex with MART-1/Melan-A

55

Molecular recognition – Van der Waals interactions



Molecular recognition – Van der Waals interactions



57

Molecular recognition – Van der Waals interactions

Do not require charges or partial charges on atoms

van der Waals interactions are considered as non-polar interactions
... even though they are electrostatic by nature

Interactions particularly **important for non-polar residues**:

- Alanine, Valine, Leucine, Isoleucine, Proline
- Cysteine, Methionine
- Phenylalanine, Tyrosine, Tryptophan

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Molecular recognition – Van der Waals interactions

Each atom tries to be positioned at optimal distance from its neighbors

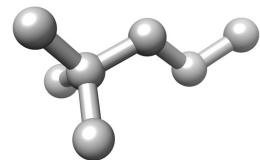
2 atoms



3 atoms



4 atoms

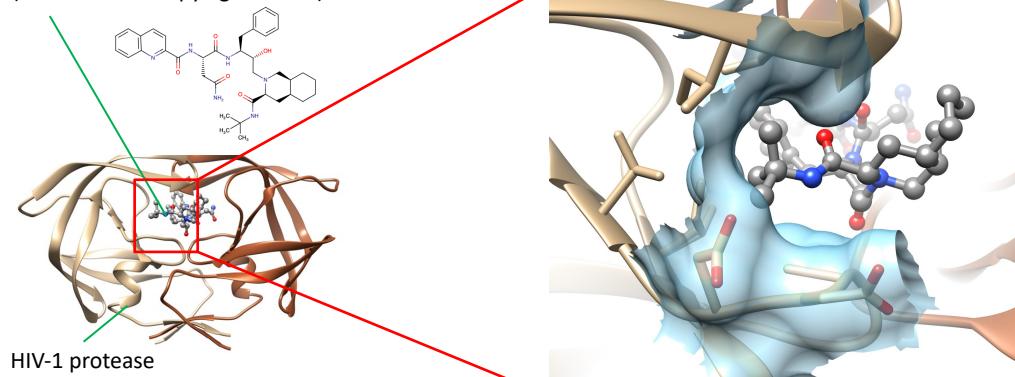


However, in molecules, atoms are also linked via covalent bonds, which force a geometry...

Molecular recognition – Van der Waals interactions

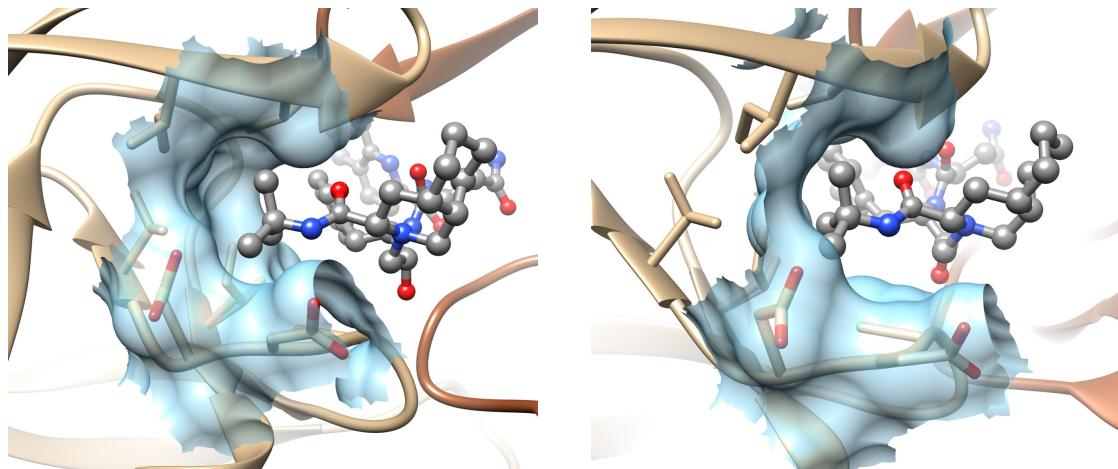
Each atom tries to be positioned at optimal distance from its neighbors

Saquinavir. HIV-1 protease inhibitor
(Used in tri-therapy against HIV)



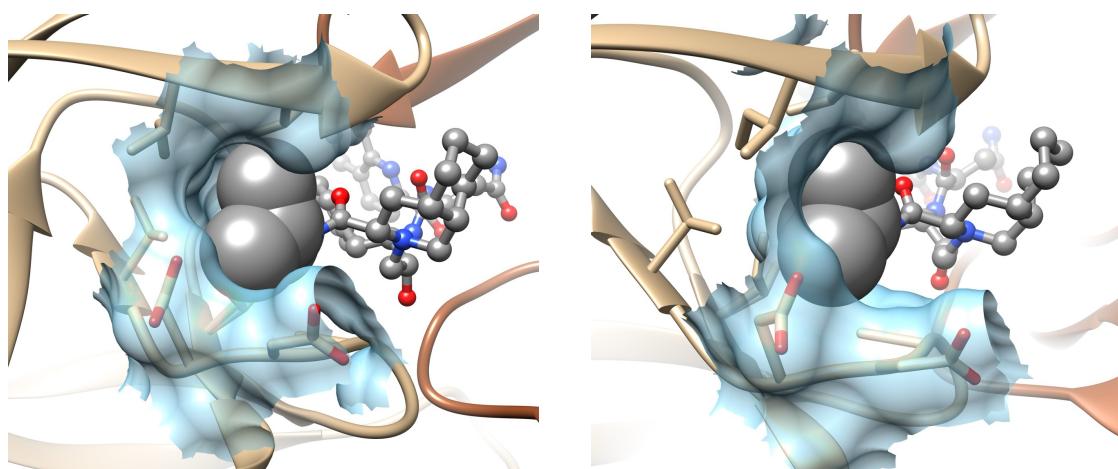
Molecular recognition – Van der Waals interactions

Each atom tries to be positioned at optimal distance from its neighbors

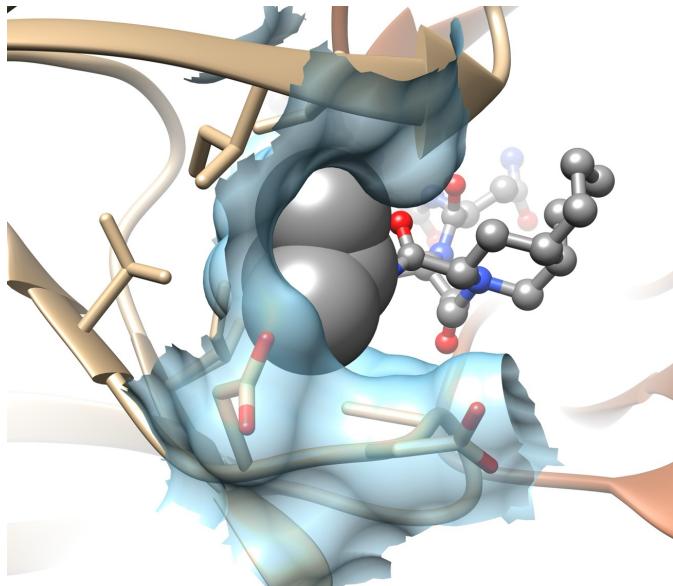


Molecular recognition – Van der Waals interactions

Each atom tries to be positioned at optimal distance from its neighbors



Molecular recognition – Van der Waals interactions



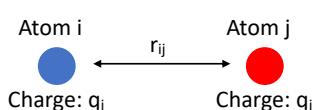
Each atom tries to be positioned at optimal distance from its neighbors

van der Waals interactions contribute therefore to:

- **packing of atoms** (and macromolecule folding)
- **shape complementarity** between binding molecules (example: protein/protein or ligand/protéine complexes)

Molecular recognition – Electrostatic interactions

The interaction between two point charges in a uniform medium is described by the **Coulomb law**



Coulomb energy

$$E_{\text{Coul}} = \frac{1}{4\pi\epsilon_0\epsilon} \frac{q_i q_j}{r_{ij}}$$

ϵ_0 : dielectric constant of vacuo

$$\frac{1}{4\pi\epsilon_0} = 332 \text{ (kcal/mol) } \text{\AA}/q_e^2$$

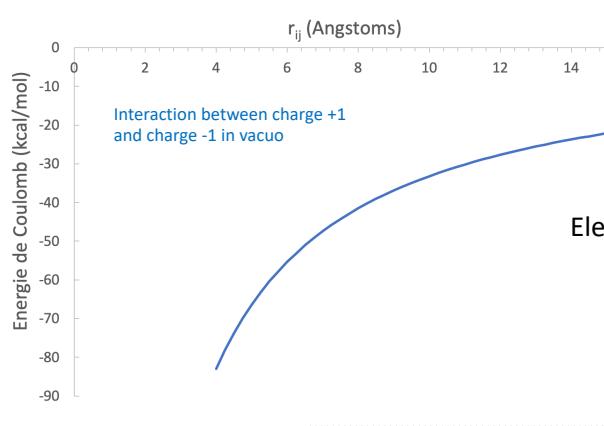
ϵ : dielectric constant of medium

$$\text{ex: } \epsilon_{(\text{vacuo})} = 1 ; \epsilon_{(\text{water})} = 80$$

Interaction between charges +1 et -1 in vacuo :

- -66 kcal/mol in vacuo

- -0.8 kcal/mol in water



Electrostatic interaction energy follows a $1/r$ expression

Long range interaction

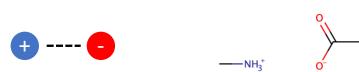
Molecular recognition – Electrostatic interactions

Electrostatic interactions can involve:

- Integer charge – integer charge

Called **ionic interactions**.

At short distance ($\sim 4/5 \text{ \AA}$), ionic interactions are called **salt bridges**.



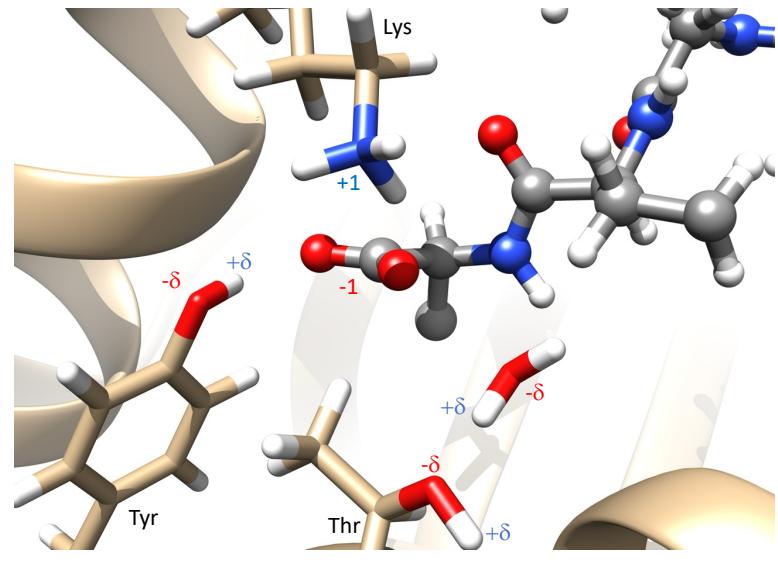
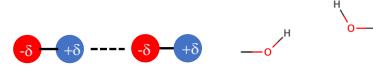
- Integer charge – permanent dipole

Ex: charged assisted hydrogen bond



- Permanent dipole – permanent dipole

Ex: hydrogen bond



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65

65

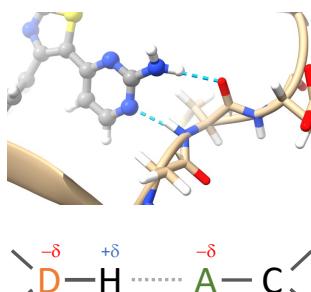
Molecular recognition – Electrostatic interactions – Hydrogen bonds

Typically between two dipoles:

- D-H where D is the hydrogen bond **donor**
- A-C where A is the hydrogen bond **acceptor** and C a carbon atom

Extremely frequent in proteins and nucleic acids

Important factor of the architecture of bio-macromolecules



Distances typiques dans les liaison hydrogène :

- Entre H et A : $\sim 1.95 \text{ \AA}$
- Entre A et D : O – O : $2.50 – 2.70 \text{ \AA}$
O – N : $2.75 – 2.85 \text{ \AA}$
N – N : $2.70 – 3.00 \text{ \AA}$

L'angle α dépend du type des atomes et de leur hybridation

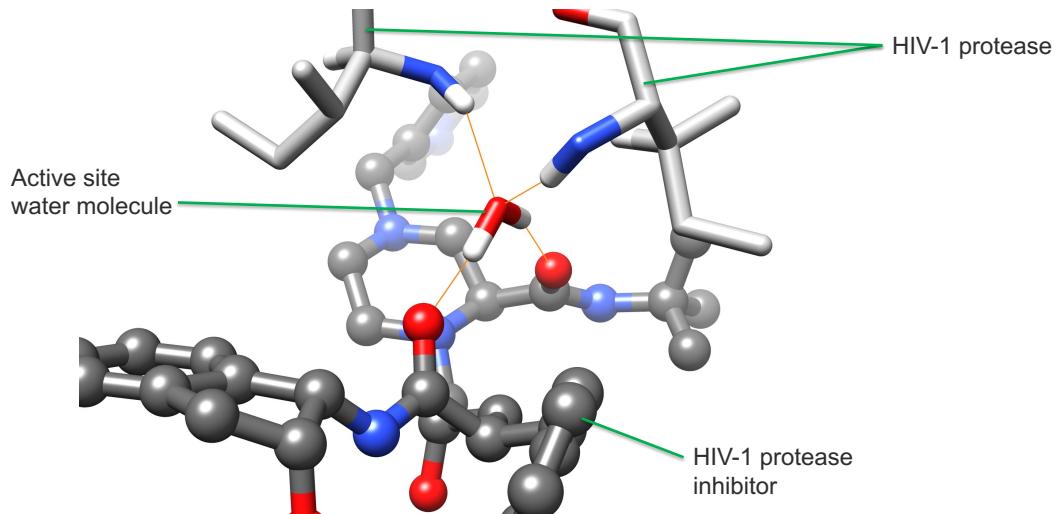
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66

66

Molecular recognition – Electrostatic interactions – Hydrogen bonds

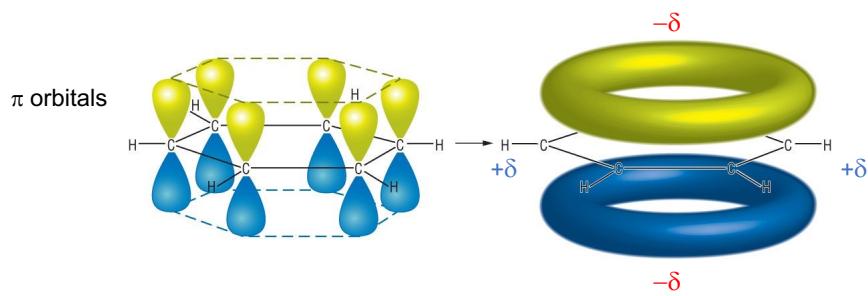


Electrostatic interactions are **local and directional** (H-bonds even more than salt bridges)

→ **Directionality / locality of interactions**
Specificity of molecular recognition

Molecular recognition – π interactions

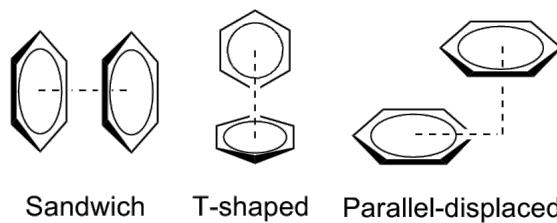
Electronic structure of benzene:



Aromatic cycles (Phenyl, Tyrosine, Tryptophan & Histidine) can interact with:

- Other aromatic cycles (stacking)
- Metals
- Polar groups
- Hydrogen bond donors

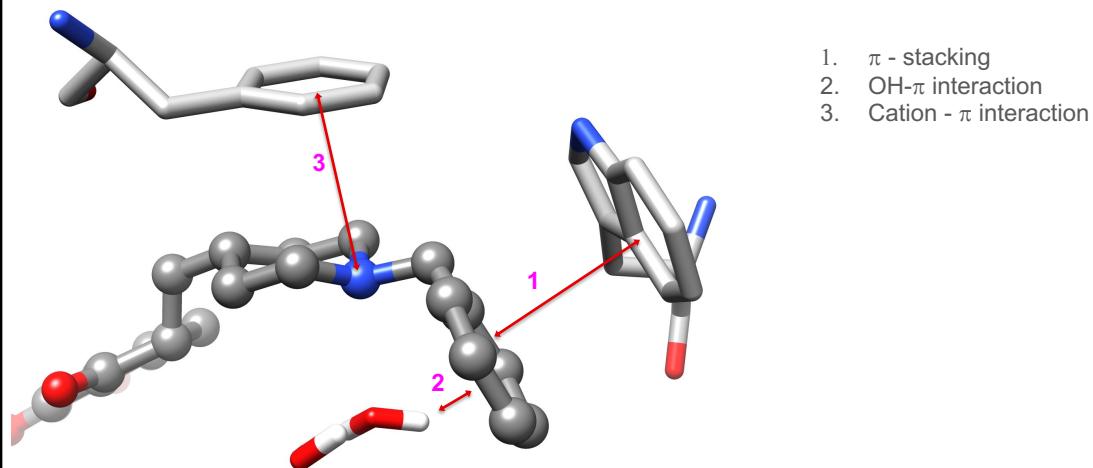
Molecular recognition – π interactions



(source: Wikipedia)

T-shaped and parallel-displaced π - π interactions are the most frequent

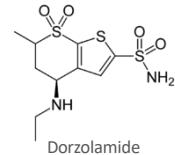
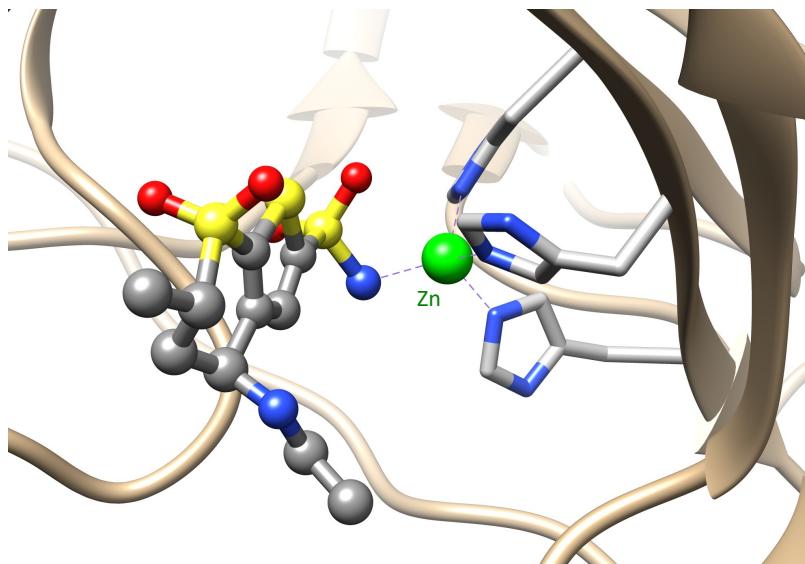
Molecular recognition – π interactions



Ex: π interactions between Donepezil and acetylcholine esterase (PDB ID 1EVE)

Molecular recognition – Metal-ion interaction

Partially covalent

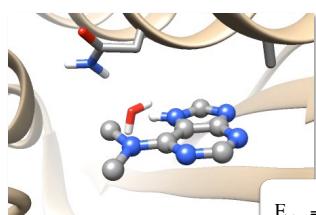


Ex: Dorzolamide, anti-glaucoma drug, in complex with carbonic anhydride II (PDB ID: 3FW3)

Molecular recognition – Other factors

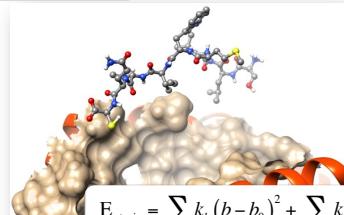
Many other factors impact the molecular recognition and binding affinity

Water bridges



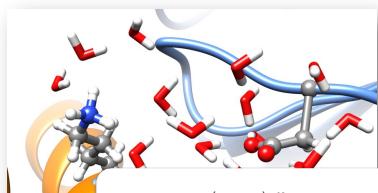
$$E_{elec} = \frac{q_i q_j}{4\pi\epsilon_0\epsilon r_{ij}}$$

Conformational changes



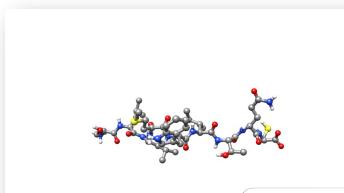
$$E_{strain} = \sum_{bonds} k_b (b - b_0)^2 + \sum_{angles} k_\theta (\theta - \theta_0)^2 + \dots$$

Desolvation and elec. shielding



$$\Delta G_{Solv} = \frac{1}{8\pi} \left(\frac{1}{\epsilon_0} - \frac{1}{\epsilon} \right) \sum_{i,j}^N \frac{q_i q_j}{\sqrt{r_{ij}^2 + a_i a_j e^{-D}}} , \quad D = \left(\frac{r_{ij}}{2\sqrt{a_i a_j}} \right)^2$$

Entropy changes

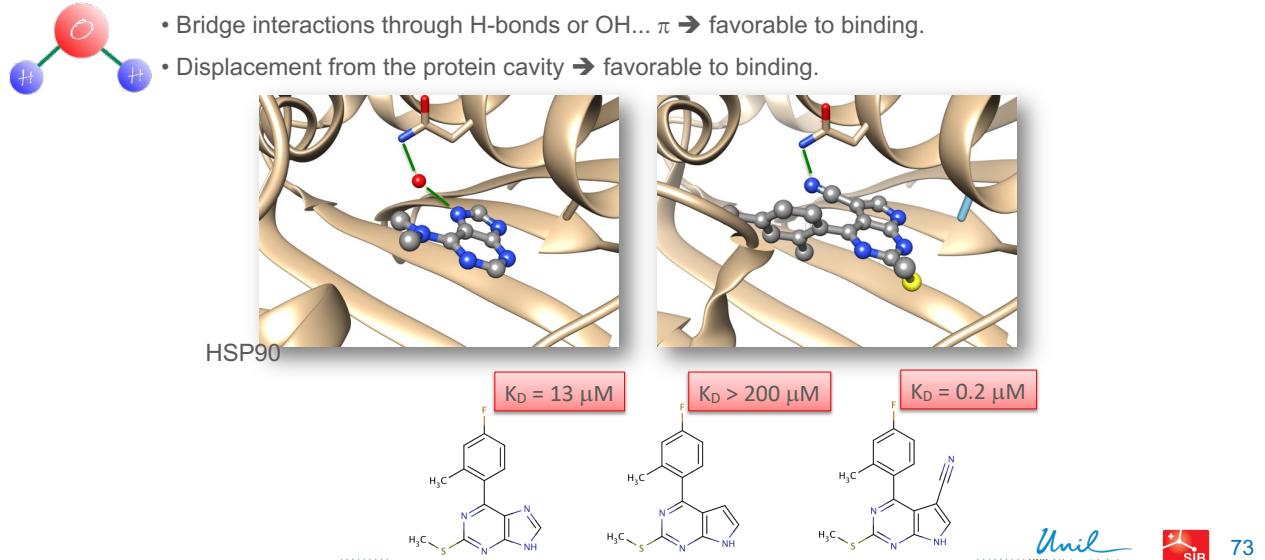


$$S = k_B \sum p_i \ln(p_i)$$

Molecular recognition – Other factors – Water

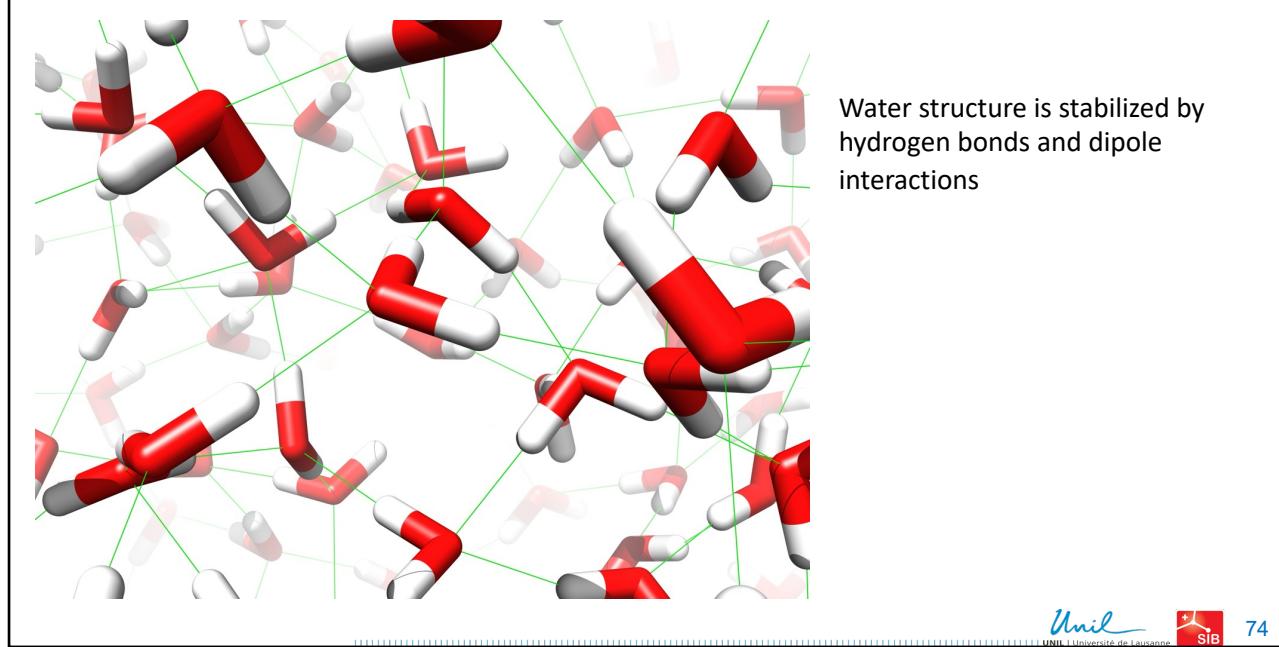
Molecular recognition between small molecule and protein takes place in an **aqueous environment**.

Discrete water molecules



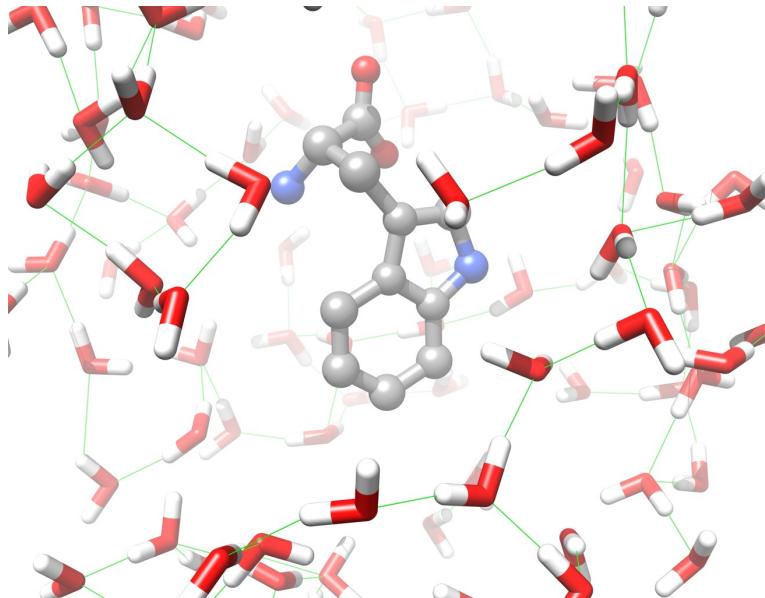
73

Molecular recognition – Other factors – Water – Hydrophobic effect



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Molecular recognition – Other factors – Water – Hydrophobic effect

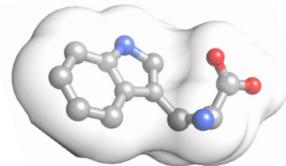


The presence of a solute decreases water-water interactions

Non-polar solvation energy is proportional to the solvent accessible surface area (SASA) for large molecules:

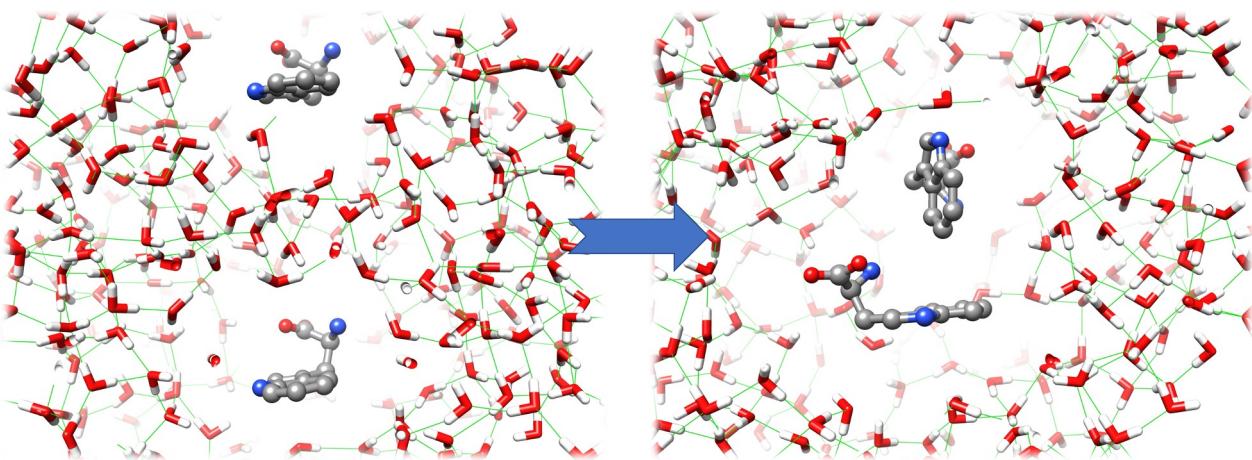
$$E = \sigma \times \text{SASA}$$

$$\sigma = 0.025 \text{ kcal}/\text{\AA}^2$$



Molecular recognition – Other factors – Water – Hydrophobic effect

Solutes aggregate to limit their deleterious on water structure

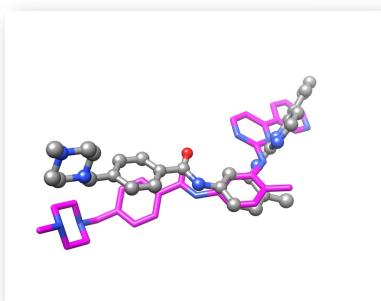
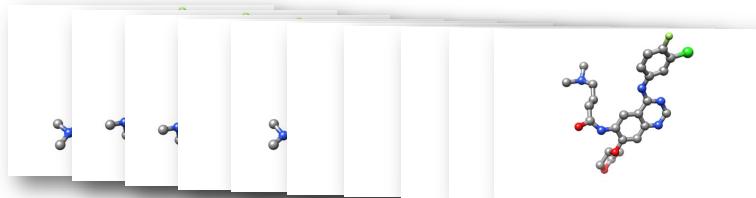


$$\text{Energy of non-polar desolvation: } \Delta G_{np} = \sigma \times \Delta \text{SASA}$$

The solvent-accessible surface area of aggregated solutes is lower than the sum of those of the separated solutes ($\Delta \text{SASA} < 0$). ΔG_{np} is therefore favorable to aggregation (binding of solutes)

Molecular recognition – Other factors – Conformational changes

Molecules have many conformations (conformers)



Ligand **bioactive conformation (geometry as bound to the protein)**

does **NOT** correspond to

Lowest energy conformation (most stable geometry in solution)

BUT is a low energy conformation (within 3 to 5 kcal/mol)

Bioactive conformation (in protein)

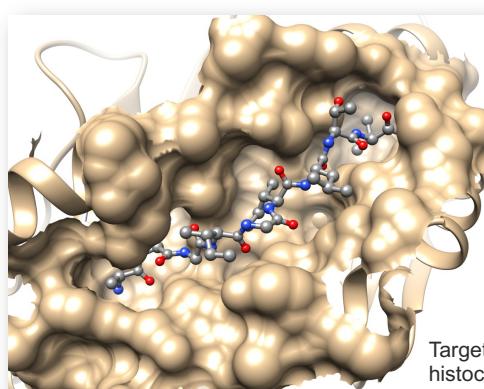
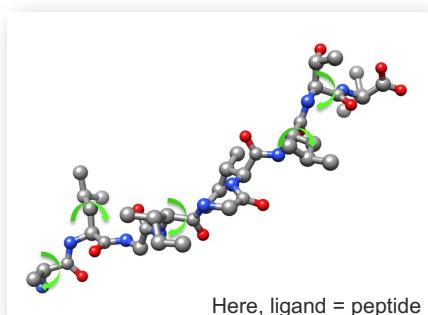
Lowest energy conformation (in solution)

Molecular recognition – Other factors – Entropy changes

Entropy is a measure of disorder. Nature likes disorder!

Loss of entropic energy when entropy (disorder) decreases.

Gain of entropic energy when entropy (disorder) increases.



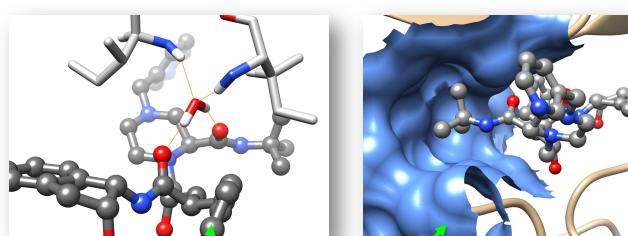
Two main events upon ligand binding to protein:

- **Conformational degrees of freedom** (rotatable bonds) are **blocked: unfavorable!**
- **Water molecules** are **kicked-out** from the protein binding site **to bulk: favorable!**

Molecular recognition – Summary

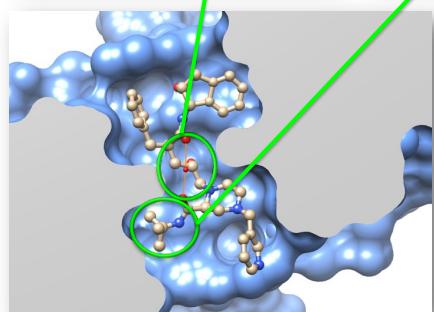
Category	Interaction	Distance	Residues involved	Remarks
Electrostatic	Ionic (charge-charge)	Long range	Arg, Lys, Asp, Glu His (if charged)	Called salt bridge at short distance
	Hydrogen bond	Short range	Arg, Lys, Asp, Glu His, Tyr Ser, Thr, Asn, Gln Cys	Directionality / locality of interactions Specificity of molecular recognition
	π interaction	Short range	Phe, Tyr, Trp, His	
Electrostatic/Non-polar	Van der Waals	Short range	Ala, Val, Ile, Leu, Pro, Cys, Met Phe, Tyr, Trp, His	Packing of atoms Shape complementarity
Non-polar	Hydrophobic effect	-	All	Solute aggregation

Molecular recognition – Potency and specificity



Various and numerous ligand-protein interactions:

- local and directional interactions
- shape complementarity



→ **Specificity**
(Limits number/nature of possible epitopes)

→ **Affinity/potency**
(Increased epitope recognition)

Molecular recognition - Molecular Motions - Molecular Dynamics



- Adding explicit droplet of water:

System solvated with explicit water molecules (TIP3P model):

- $\sim 29,500$ water molecules
- $\sim 100,000$ atoms in total

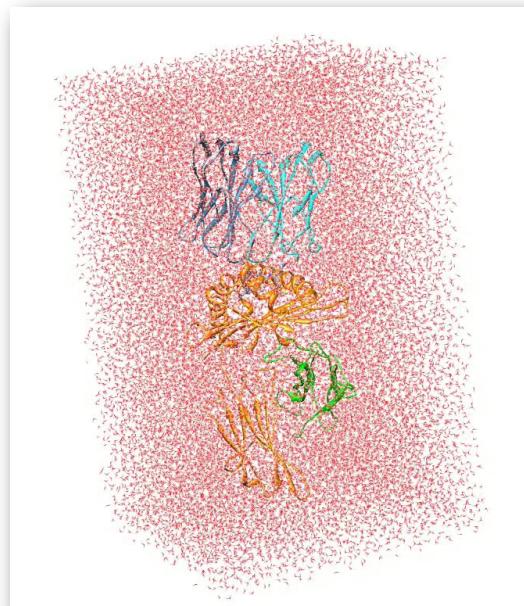
- Molecular Dynamics (MD)

Atom motions are calculated to follow Newton's equation of motion, at **300 K** and **1 atm**.

Typical simulation times: from **0.5 ns** to ~ 1000 ns (1 ns = 10^{-9} s).

→ Simulation closer to physiological reality, but more computationally intensive

Molecular recognition - Molecular Motions - Molecular Dynamics



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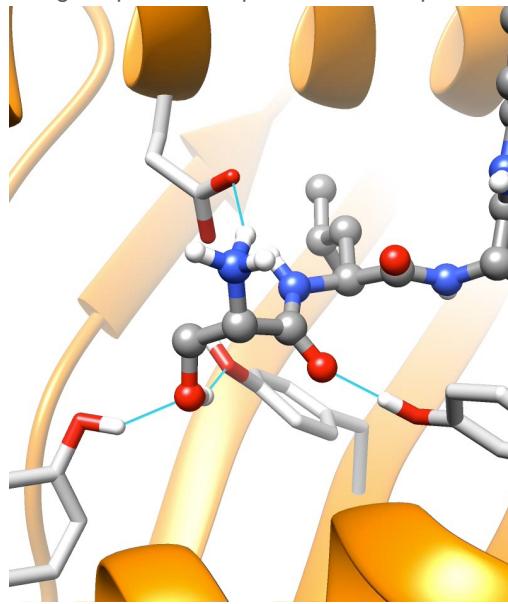
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Molecular recognition - Molecular Motions - Molecular Dynamics

Typical motions in a ligand/protein complex at room temperature:

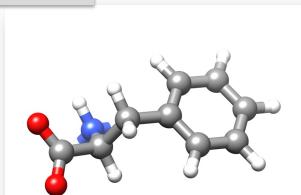


Peptide epitope in ball and stick representation

MHC protein in ribbon representation with some side chains in stick representation

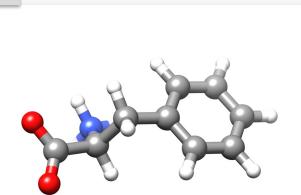
Molecular recognition – introduction to molecular mechanics

Bond length



$$E_{bond} = k_b(b - b_0)^2$$

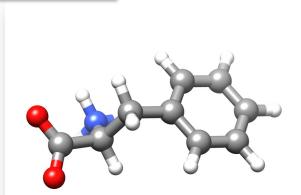
Bond angle



$$E_{angle} = k_\theta(\theta - \theta_0)^2$$

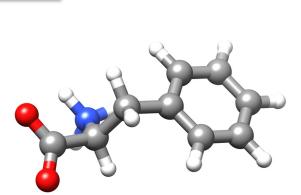
Molecular dynamics is decomposed into elementary motions

Dihedral angle



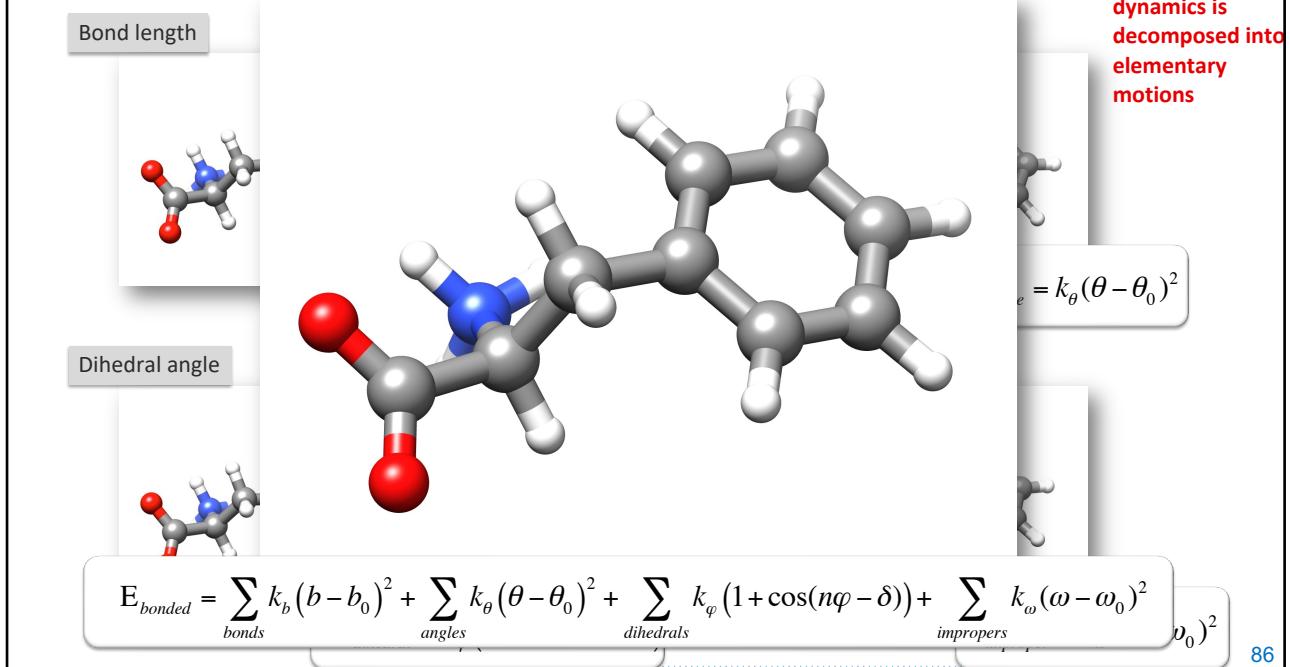
$$E_{dihedral} = k_\varphi (1 + \cos(n\varphi - \delta))$$

Improper angle



$$E_{improper} = k_\omega (\omega - \omega_0)^2$$

Molecular recognition – introduction to molecular mechanics

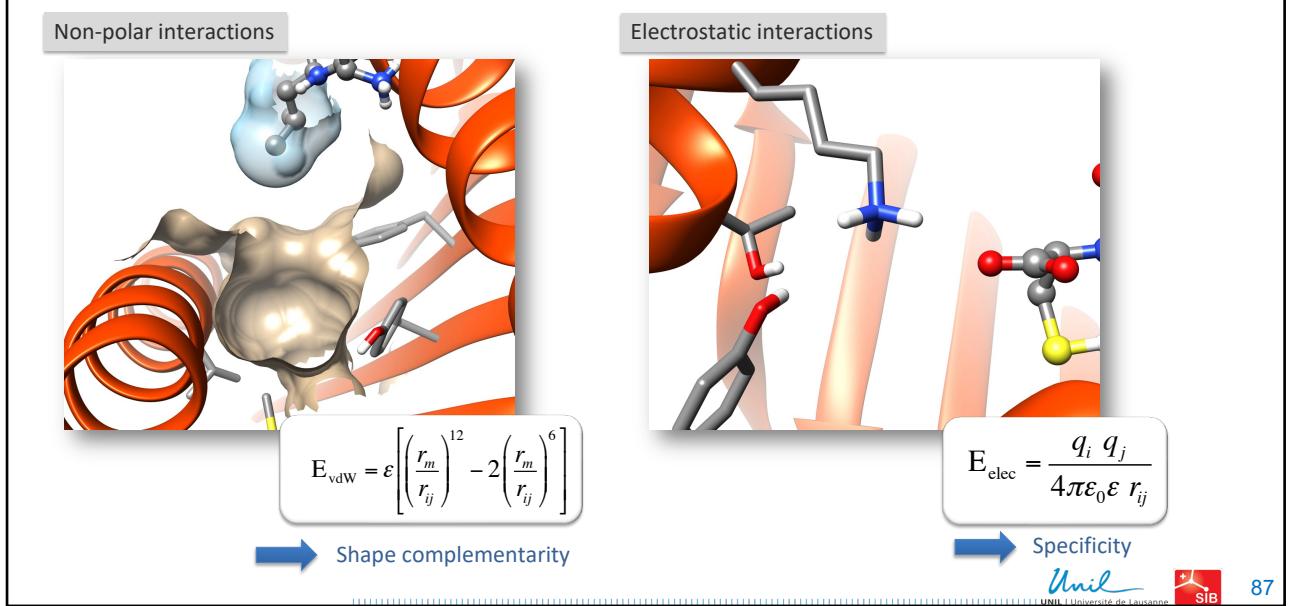


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Molecular recognition – Molecular interactions

Molecular recognition is driven by non-polar and electrostatic interactions



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Molecular recognition - type of interactions

Non Polar:

Ala, Val, Leu, Ile, Pro, Met, ~Cys

Polar:

Ser, Thr, Asn, Gln, Tyr, His, Trp, ~Cys

Aromatic:

Phe, Tyr, Trp, His

Negatively charged:

Asp, Glu

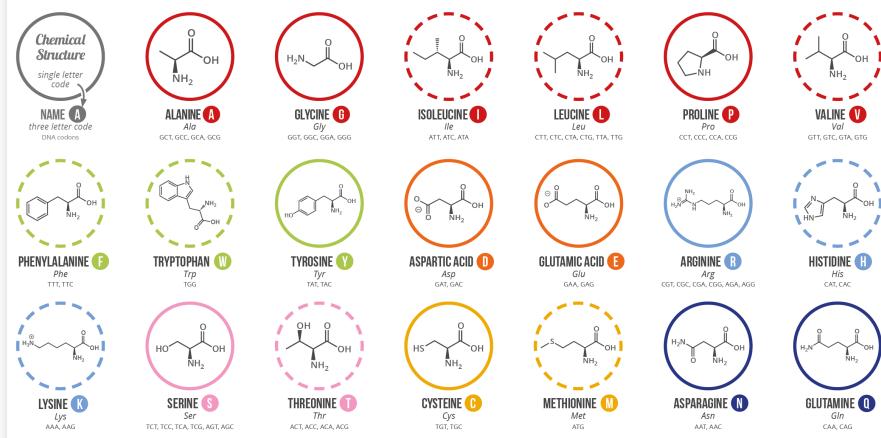
Positively charged:

Arg, Lys, ~His

A GUIDE TO THE TWENTY COMMON AMINO ACIDS

AMINO ACIDS ARE THE BUILDING BLOCKS OF PROTEINS IN LIVING ORGANISMS. THERE ARE OVER 500 AMINO ACIDS FOUND IN NATURE - HOWEVER, THE HUMAN GENETIC CODE ONLY DIRECTLY ENCODES 20. 'ESSENTIAL' AMINO ACIDS MUST BE OBTAINED FROM THE DIET, WHILST NON-ESSENTIAL AMINO ACIDS CAN BE SYNTHESISED IN THE BODY.

Chart Key: ● ALIPHATIC ● AROMATIC ● ACIDIC ● BASIC ● HYDROXYLIC ● SULFUR-CONTAINING ● AMIDIC ● ○ NON-ESSENTIAL ● ○ ESSENTIAL



Note: This chart only shows those amino acids for which the human genetic code directly codes for. Selenocysteine is often referred to as the 21st amino acid, but is encoded in a special manner. In some cases, distinguishing between asparagine/aspartic acid and glutamine/glutamic acid is difficult. In these cases, the codes asx (B) and glx (Z) are respectively used.

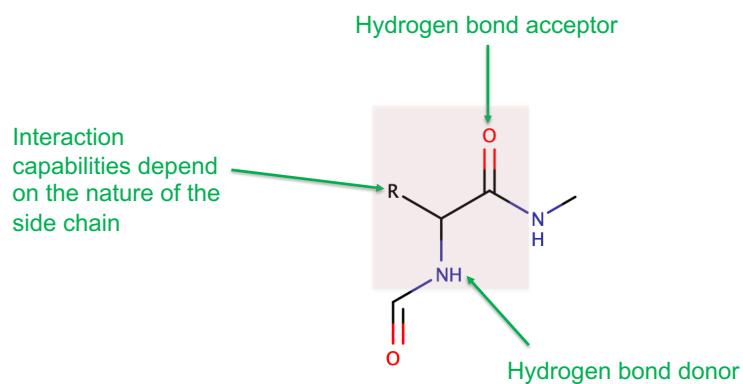
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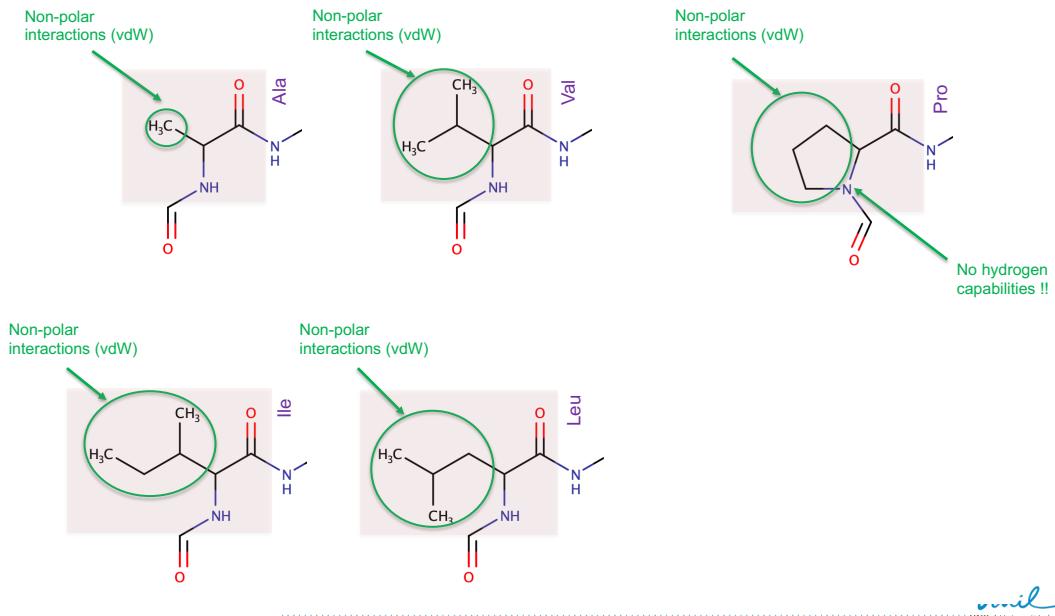
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Interactions Moléculaires – Backbone des acides aminés



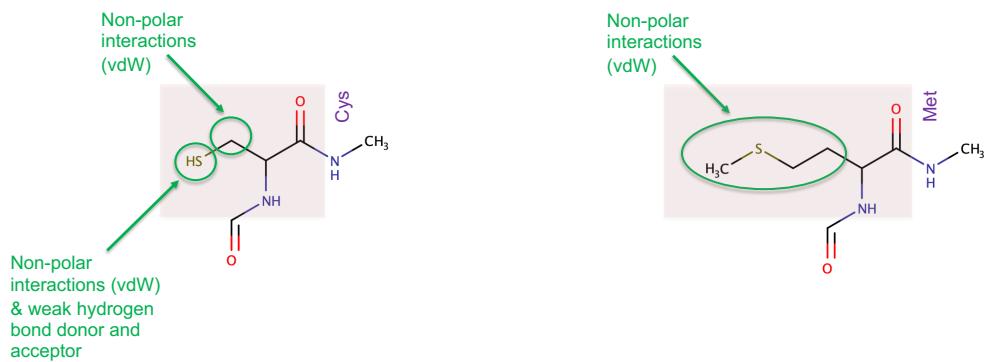
89

Interactions Moléculaires – Chaînes latérales des acides aminés



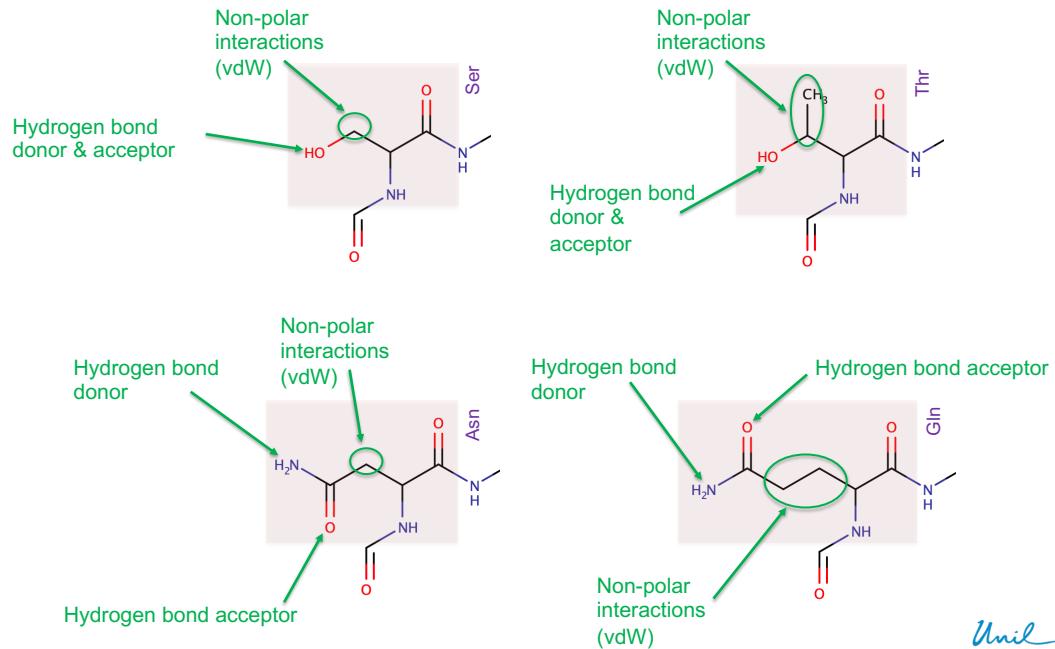
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Interactions Moléculaires – Chaînes latérales des acides aminés

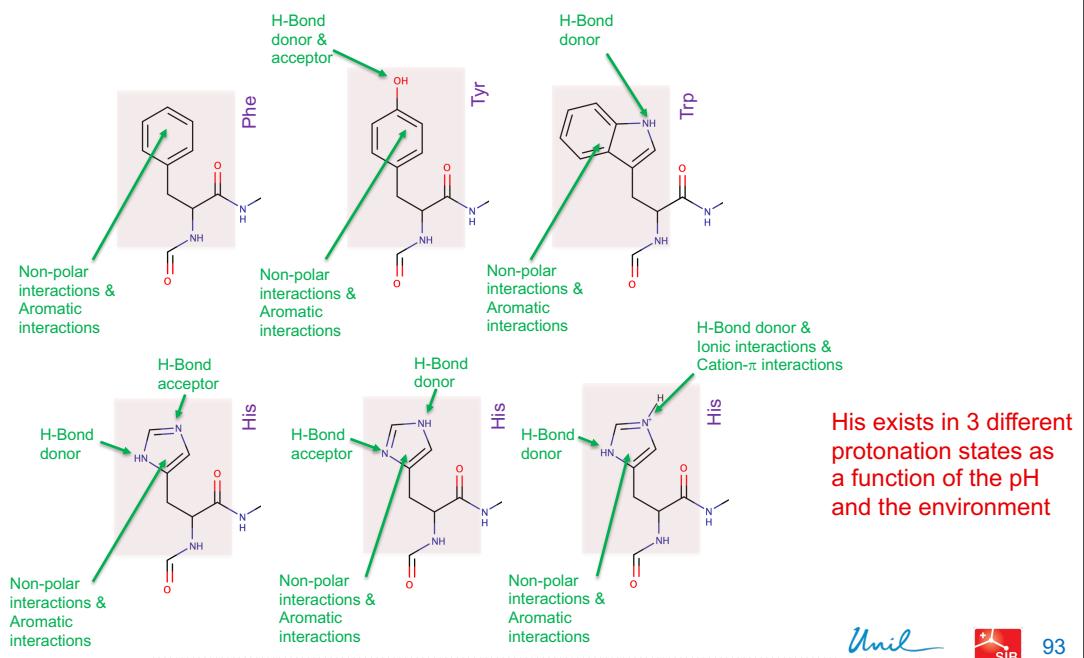


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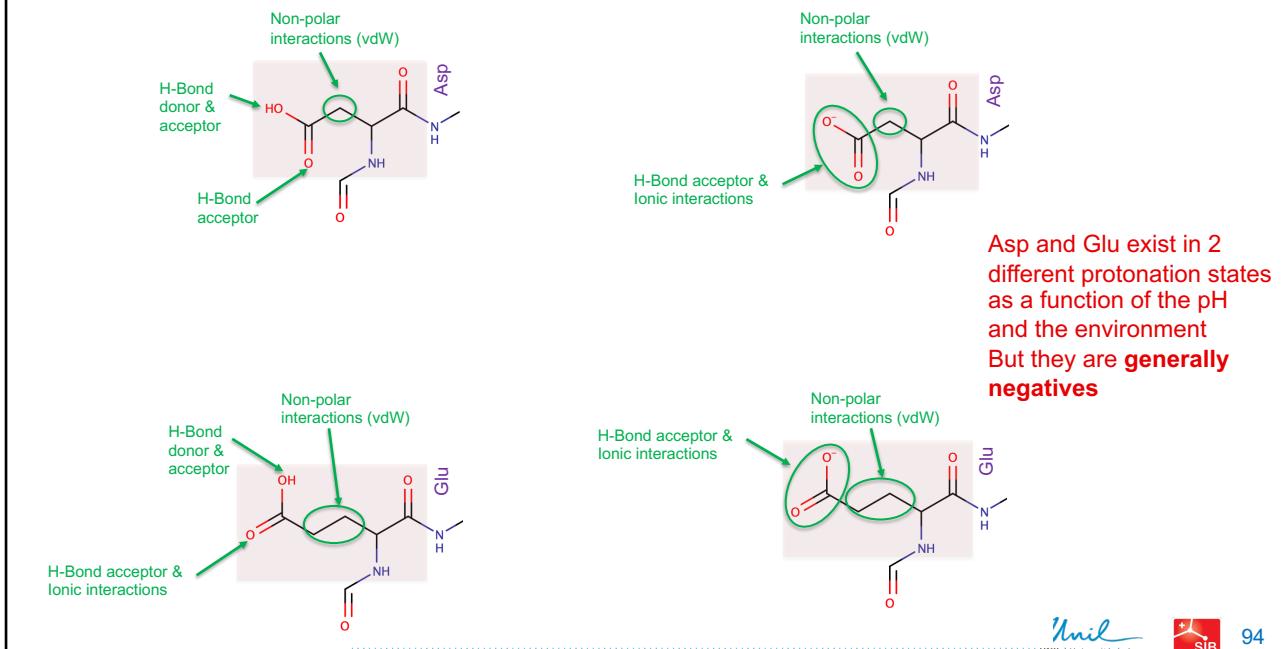
Interactions Moléculaires – Chaînes latérales des acides aminés



Interactions Moléculaires – Chaînes latérales des acides aminés

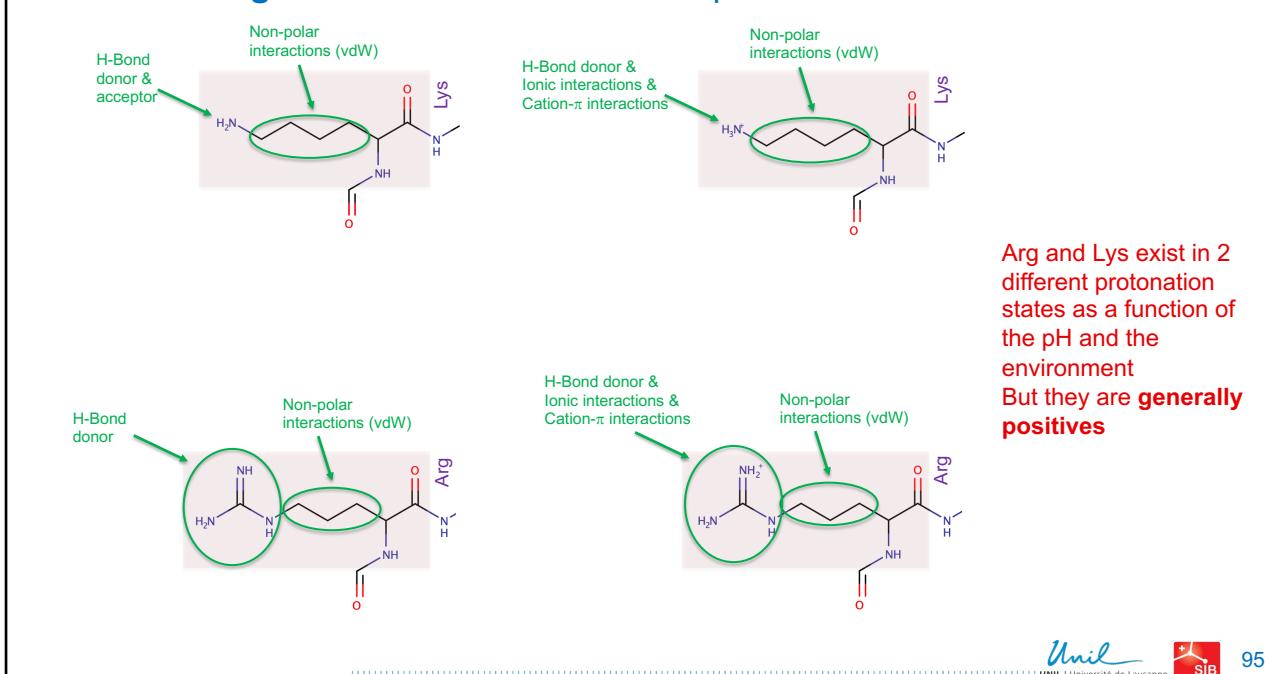


Molecular recognition – Possible interactions per amino acids



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Molecular recognition – Possible interactions per amino acids



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Molecular recognition

Let's start with UCSF Chimera !!!

Contacts: vincent.zoete@unil.ch , antoine.daina@sib.swiss